## COLONIZATION OF THE OASIS MALARIA VECTOR, ANOPHELES SERGENTII, IN EGYPT<sup>1</sup>

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Anopheles sergentii (Theobald) is the principal vector of oasis-desert malaria, and shows unique physiological and behavioral adaptability in its extreme environment (Farid 1956). Throughout the range of this species, including North Africa, the Middle East and Pakistan (Knight and Stone 1977), it is an efficient malaria vector (Farid 1956). In Egypt, An. sergentii transmits malaria in the Western Desert Oases and in Faiyum Oasis (Halawani and Shawarby 1957), and on two occasions has been found naturally infected with sporozoites (Farid 1940; J. C. Beier and M. A. Kenawy, unpublished data).

Studies on An. sergentii in the laboratory have been limited to observations of field-collected specimens (Saliternik 1955) due to the lack of laboratory colonies. Many unsuccessful efforts have been made to colonize this species in Egypt (Farid 1956; M. A. Kenawy, personal communication). Except for an unpublished report to WHO on the colonization of this species from Jordan by G. Davidson in 1967 (see Zahar<sup>4</sup>), we are unaware of other colonization attempts. The main obstacle to colonization is that this species is eurygamous as it does not mate readily under laboratory conditions. This report describes the establishment of three colonies of An. sergentii.

The initial An. sergentii colony originated from more than 500 larvae collected in Siwa Oasis, Matrouh Governorate, Egypt, during October 1982. The other two colonies originated from collections of over 200 larvae from Sinnuris District, Faiyum Governorate during June and October, 1983. Using artificial mating

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<sup>4</sup> Zahar, A. R. 1973. Review of the ecology of malaria vectors in the WHO Eastern Mediterranean Region. WHO/MAL/73.808 21 p.

techniques, the Siwa colony was maintained nine months through the eighth generation. The Faiyum strains were maintained for 10 generations over one year.

Field collected larvae were brought to the insectary in Cairo for colonization. The insectary was maintained at 20–30°C, 40–80% RH, with illumination by fluorescent lighting for approximately 8 hours a day.

The artificial mating technique was similar to that employed at the National Institutes of Health (R. Gwadz, personal communication) and described by Ow Yang et al. (1963). Females, held either in a 30 cm<sup>3</sup> cage, or in a 300 ml screened cup, were offered a human blood meal immediately prior to mating. Males were lightly anesthetized by ether and pinned to an applicator stick with a minuten pin. This species, in comparison to other established Anopheles colonies maintained by this technique was difficult to force-mate and all matings were made using a dissecting microscope at 100-150X. Experienced individuals could mate up to 20 females per hour. Matings were most successful using males 3-6 days of age.

Mated females were held in 30 cm<sup>3</sup> wooden framed cages and provided with 10% sucrose on cotton which was changed daily. A 12 cm diam. bowl lined with filter paper and 34 filled with distilled water was used for oviposition. Eggs were collected and usually allowed to hatch in the oviposition bowls. Mated females were refed in the cages on human blood and it was common to refeed the same females up to 6 times over a 1-month period, as most mated females survived 1 month.

First instar larvae were placed in 30 cm diam. round enameled pans half-filled with water. Usually, larvae were reared in a mud slurry (2 kg clay soil stirred in 50 liters tap water; water was used for larvae after 2 days). Preliminary experiments showed faster development and better survival in this water when compared to distilled or tap water. Larvae were fed ground dog food pellets (fat-free), once a day.

Observations were made using the Faiyum colonies in generations 2 to 6 held at  $27 \pm 2^{\circ}$ C. With individual females held in 300 ml screened paper cups, containing 50 ml of water; the duration of the first and second gonotrophic cycles were  $4.9 \pm 0.9$  (n = 55) and  $3.6 \pm 0.7$  (n = 11), days, respectively. The number of eggs per female ranged from 21 to 148, with means of  $66.4 \pm 26.8$  and  $74.1 \pm 23.7$  for the first and second gonotrophic cycles, respectively. Usually, eggs hatched in 2 days and the mean percentage hatch was 81% for 14 replicates containing at least 50 eggs. In larval rearing experiments with 10 replicates of 50 larvae per pan in mud slurry preparation, the

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mean pupation time was 10.4 days, with 81% survival. For 10 replicates using tap water, and 8 replicates of distilled water, mean pupation times were 13.2 days and 15.2 days, with mean survival rates of 55% and 33%, respectively. Although adult females were usually fed on human blood, they also readily fed on pigeons, mice and guinea pigs.

Natural mating in cages was not observed, and on several occasions females were dissected and found to be uninseminated, including those from trials with large cages (90 cm<sup>3</sup>) containing over 500 adults. Artificial mating appears to be necessary for colony maintenance. However, as the survival and parity potential of the species is high under laboratory conditions, the number of females mated per generation does not have to be great for routine colony maintenance.

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