ROLE OF ANOPHELES CULICIFACIES S.L. AND AN. PULCHERRIMUS IN MALARIA TRANSMISSION IN GHASSREGHAND (BALUCHISTAN), IRAN¹

M. ZAIM,² S. K. SUBBARAO,³ A. V. MANOUCHEHRI² AND A. H. COCHRANE⁴

ABSTRACT. A 2-site immunoradiometric assay (IRMA) was performed on the head and thorax of Anopheles culicifacies s.l. and An. pulcherrimus females, the 2 most common anopheline species in the District of Ghassreghand (Baluchistan, Iran), collected during the 2 peak malaria transmission seasons (May and September-October 1991). Positive IRMA results revealed the 2 species as potential vectors of malaria in this highly endemic district. This finding serves as the first report on natural infection of An. pulcherrimus in Iran and is the second on natural infection of An. culicifacies since the previous report of 1959.

INTRODUCTION

Malaria is one of the major health problems in Baluchistan, southeastern Iran from which, in recent years more than 50% of the country's total cases have been reported. In 1991 with about 41,000 reported cases of malaria (annual parasite incidence of 28 in 1,000 population), this area has been confronted with the highest number of malaria cases in the past 16 years (Centre for Disease Control 1991).

In recent years, more than 80% of the cases of malaria in Baluchistan has been reported from the "Shahrestans" of Iranshahr and Chabahar where Anopheles culicifacies sensu lato is the most common anopheline species. This species has been incriminated as a vector of malaria in Iran during an epidemic outbreak in Zabol (Sistan and Baluchistan Province) in 1959 (Manouchehri and Ghiassedin 1959). Also in these 2 named shahrestans, four other known malaria vectors of Iran exist; however, their role in transmission in this part of the country has not been studied. These latter species are An. dthali Patton, An. fluviatilis James, An. stephensi Liston and An. superpictus Grassi (Zaim 1987). Anopheles pulcherrimus Theobald, which has been incriminated as a vector of malaria in Iraq on epidemiological grounds (Rishikesh 1972) and in central Asia (Christophers 1933) and Kunduz area, northeast Afghanistan through natural infection (M. S. Badaway, in Zahar 1974), is also present in relatively high numbers in the rice growing regions of Chabahar (districts of Ghassreghand and Nikshahr, 300–900 m above sea level) (Zaim et al. 1992).

A highly sensitive immunodiagnostic technique, a 2-site immunoradiometric assay (IRMA) based on species-specific monoclonal antibodies (mAbs) has been developed by Zavala et al. (1982) that can detect, identify and quantify sporozoites in individual or pools of mosquitoes. The assay has been successfully used for the determination of the *Plasmodium vivax* and *P. falciparum* sporozoite rate and load in fresh and dried field collected mosquitoes (Collins et al. 1984, Subbarao et al. 1988).

This study uses the IRMA to examine the potential role of An. culicifacies s.l. and An. pulcherrimus, the 2 most common anophelines in the Ghassreghand (Baluchistan) where the annual parasite incidence of malaria averages about 60/1,000 population.

MATERIALS AND METHODS

Study area: The investigation was carried out over a period of 11 months (February–December 1991) in the District of Ghassreghand, Shahrestan of Chabahar. Streams and stream fed pools, rice fields and palm irrigation plots are the main sources of mosquito breeding in the area. The average maximum and minimum temperatures in summer are 39.7° and 26.2°C and in winter 24.1° and 11.8°C, respectively. The average annual rainfall is about 100 mm. The district has been under an indoor residual spraying program for malaria control using malathion (50% wp, 2 g/m²) in February and primiphos-methyl (Actellic) (40% wp, 2 g/m²) in September 1991.

Mosquito collections: Pyrethrum space spray catches were performed, once a month in 4 villages (Bocan, Hit, Homeiry and Zeineddini). In each village 8 fixed shelters (4 human and 4 animal shelters) were sampled. Mosquitoes were identified to species and the collections during the 2 main peaks of malaria transmission, i.e.,

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² Department of Medical Entomology, School of Public Health, P.O. Box 6446, Teheran 14155, Iran.

³ Malaria Research Centre, 22 Sham Nath Marg, Delhi 110 054, India.

⁴ Department of Medical and Molecular Parasitology, New York University School of Medicine, 550 First Ave., New York, NY, USA.

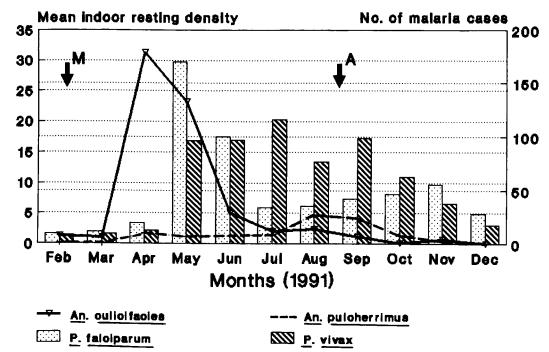


Fig. 1. Monthly indoor resting densities of Anopheles culicifacies s.l. and An. pulcherrimus and the number of malaria cases reported in Ghassreghand (Baluchistan, Iran), February–December 1991. M and A indicate the months the district was sprayed with 2 g/m^2 malathion and primiphos-methyl (Actellic), respectively.

May and September-October, were dried at room temperature and stored in a refrigerator, for use in IRMA, after their abdomens were cut off.

Two-site IRMA: Mosquitoes were assayed for Plasmodium infections using a previously described 2-site IRMA (Zavala et al. 1982, 1983), with the following modifications: pools of 10 mosquitoes were homogenized in $100~\mu l$ grinding buffer and $300~\mu l$ phosphate buffer saline-10% bovine serum albumin was added to make a final volume of $400~\mu l$. Thirty μl of this homogenate was used as antigen source. The remaining procedure was the same as described by Subbarao et al. (1988).

Positive controls consisting of extracts of *P. vivax* and *P. falciparum* sporozoites and negative controls consisting of extracts of mosquitoes reared in the laboratory, were included in each microtiter plate. Unknown preparations giving readings at least twice as high as the maximum negative control value were considered positive. Suspected positive extracts were reassayed to confirm their positivity.

MAbs 2F2 and 2A10, which recognize a species-specific repetitive epitope on the circumsporozoite protein of *P. vivax* and *P. falciparum*, respectively, were used (Nardin et al. 1982). The

mAbs were labeled with I125 (Amsterdam Laboratories, Arlington Heights, IL).

RESULTS AND DISCUSSION

The mean indoor resting densities of An. culicifacies s.l. and An. pulcherrimus and the number of reported malaria cases in the district of Ghassreghand are presented in Fig. 1. Both species were active throughout most of the year and the peak indoor resting density, under the ongoing malaria control measures, was found to be in April and August. A total of 1,214 malaria cases were reported in this area during the study period, with the highest numbers reported in May and September (Fig. 1).

Table 1 summarizes the results of the IRMA performed on An. culicifacies s.l., An. pulcherrimus and the 3 other anophelines (An. dthali, An. fluviatilis and An. stephensi) collected in indoor resting sites during the peak malaria transmission seasons in Ghassreghand. Out of the 2,305 mosquitoes collected during May 1991 and tested for Plasmodium infection, 85% and 10.7% belonged to An. culicifacies s.l. and An. pulcherrimus, respectively, of which 2 pools of An. culicifacies s.l. were found positive for P. vivax. Also out of the 1,245 females collected

Table 1. Results of the immunoradiometric assay performed on pools of 10 mosquitoes collected during peak malaria transmission seasons in Ghassreghand, Baluchistan, Iran (1991).

Species	May 1991		SeptOct. 1991	
	No. tested	No. of positive pools*	No.	No. of positive pools*
An. culicifacies	1,960	2	276	
An. dthali	26	_	11	_
An. fluviatilis	56		7	_
An. pulcherri- mus	246	_	686	2
An. stephensi	17	_	265	_

^{*} Only positive for P. vivax.

during September-October 1991, An. culicifacies s.l. and An. pulcherrimus constituted 22.2% and 55.1% of the collected material, respectively, of which 2 pools of An. pulcherrimus were positive for P. vivax. Surprisingly no positives were found for P. falciparum in this study.

Our recent study on the seasonal activity of An. culicifacies s.l. and An. pulcherrimus in a village where no anti-mosquito control measures have been in practice, has revealed that An. culicifacies s.l. has 2 peaks of activity in the district of Ghassreghand, one in April-May (main peak) and the other in October-November (Zaim et al., unpublished data). Anopheles pulcherrimus has also 2 peaks of activity, one in April-May and the other in August-September (Zaim et al. 1992). Because the second peak for malaria cases in the district of Ghassreghand is in September, An. pulcherrimus has been suspected as a vector in this area. The present IRMA results correspond well with the observed vector population and malaria cases in the study area and serves as the first report on natural infection of An. pulcherrimus in Iran and is the second report on natural infection of An. culicifacies s.l. in Sistan and Baluchistan province since the previous report of 1959. It is noteworthy that in preliminary studies carried out in Iran only species A of An. culicifacies has been found (Zaim and Javaherian 1991); however, the study is still in progress.

The distribution of An. pulcherrimus is mainly restricted to the rice growing areas of the Iranshahr and Chabahar, i.e., districts of Ghassreghand and Nikshahr. Based on epidemiological data, An. culicifacies s.l. is considered as the primary vector of malaria in the other areas of the 2 highly endemic shahrestans. Anopheles stephensi, which used to be a prevalent species in this part of the country, has responded well

to indoor residual spraying programs, and during 1990–91 it comprised less than 4% of the pyrethrum space spray catches of human and animal shelters in the 2 named shahrestans (Zaim, unpublished data). Since positive IRMA results on potential vectors do not prove vector incrimination, the determination of the natural infection rates of An. culicifacies and An. pulcherrimus based on salivary gland dissections and comparative vector potential studies that are currently underway should bring more light to the relative importance of these 2 species in Baluchistan, Iran.

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REFERENCES CITED

Centre for Disease Control. 1991. Annual report on Malaria and Bilharziasis control in Iran. 14 pp. (In Persian).

Christophers, S. R. 1933. The fauna of British India, including Ceylon and Burma. Diptera. Vol. 4. Family Culicidae. Tribe Anophelini. Taylor and Francis, London.

Collins, F. H., F. Zavala, P. M. Graves, A. H. Cochrane, R. W. Gwadz, J. Akoh and R. S. Nussenzweig. 1984. First field trial of an immunoradiometric assay for the detection of malaria sporozoites in mosquitoes. Am. J. Trop. Med. Hyg. 33:538-543.

Manouchehri, A. V. and M. Ghiassedin. 1959. Inst. Prasitol. Malariol., Annual Report, Teheran, Iran. Publ. 669. (In Persian).

Nardin, E. H., V. Nussenzweig, R. S. Nussenzweig, W. E. Collins, K. T. Harinasuta, P. Tapchaisri and Y. Chomcharn. 1982. Circumsporozoite proteins of human malaria parasites P. falciparum and P. vivax. J. Exp. Med. 156:20–30.

Rishikesh, N. 1972. Anopheles pulcherrimus Theobald (1902) as probable vector of malaria in Iraq. Bull. Endem. Dis. 13:7-13.

Subbarao, S. K., T. Adak, K. Vasantha, H. Joshi, K. Raghavendra, A. H. Cochrane, R. S. Nussenzweig and V. P. Sharma. 1988. Susceptibility of Anopheles culicifacies species A and B to Plasmodium vivax and Plasmodium falciparum as determined by immunoradiometric assay. Trans. R. Soc. Trop. Med. Hyg. 82:394-397.

Zahar, A. R. 1974. Review of the ecology of malaria vectors in the WHO Eastern Mediterranean Region. Bull. W.H.O. 50:427-440.

Zaim, M. 1987. Malaria control in Iran-present and future. J. Am. Mosq. Control Assoc. 3:392-396.

- Zaim, M. and Z. Javaherian. 1991. Occurrence of Anopheles culicifacies species A in Iran. J. Am. Mosq. Control Assoc. 7:324-326.
- Zaim, M., A. V. Manouchehri, M. Motabar, H. Ladonni, G. Mowlaii, M. H. Kayedi, K. Pakdad and M. Nazari. 1992. Ecology of Anopheles pulcherrimus in Baluchistan, Iran. J. Am. Mosq. Control Assoc. 8:293-296.
- Zavala, F., R. W. Gwadz, F. H. Collins, R. S. Nussen-
- zweig and V. Nussenzweig. 1982. Monoclonal antibodies to circumsporozoite proteins identify the species of malaria parasite in infected mosquitoes. Nature 299:737–738.
- Zavala, F., A. H. Cochrane, E. H. Nardin, R. S. Nussenzweig and V. Nussenzweig. 1983. Circumsporozoite proteins of malaria parasites contain a single immunodominant region with two or more identical epitopes. J. Exp. Med. 157:1947-1957.