INDOOR THERMAL FOGGING AGAINST VECTOR MOSQUITOES WITH TWO BACILLUS THURINGIENSIS ISRAELENSIS FORMULATIONS, VECTOBAC ABG 6511 WATER-DISPERSIBLE GRANULES AND VECTOBAC 12AS® LIQUID

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ABSTRACT. Bioefficacy of thermal fogging application of 2 *Bacillus thuringiensis israelensis* formulations, Vectobac ABG 6511 water-dispersible granules (3,000 international toxic unit [ITU]/mg) and Vectobac 12AS[®] liquid (1,200 ITU/mg), was assessed for larvicidal activities against *Aedes aegypti*, *Aedes albopictus*, *Anopheles dirus*, and *Culex quinquefasciatus*. Portable Agrofog[®] AF35 sprayers were used to apply the 2 formulations indoors in residential premises on Penang Island, Malaysia. Vectobac ABG 6511 showed good larvicidal effect against all 4 mosquito species at 3 of the higher doses tested (2.91 × 10[°], 1.45 × 10[°], and 0.71 × 10[°] ITU/ha), with more than 96% mortality at 48 h after spraying. As a comparative formulation, Vectobac 12AS also showed good larvicidal activity against all 4 mosquito species at 2 of the higher doses tested (2.87 × 10[°] and 1.46 × 10[°] ITU/ha), with more than 92.5% mortality at 48 h after spraying. Larvae of *An. dirus* were significantly more susceptible to both water-based Vectobac formulations when compared to the other 3 mosquito species. Both microbial formulations showed better efficacy at higher doses. However, even at the lowest dose tested, Vectobac ABG 6511 and Vectobac 12AS (both at 0.36 × 10[°] ITU/ha) showed larvicidal properties, with more than 66% mortality at 48 h after spraying. Overall, for this bacterial agent, the water-dispersible granule formulation has better prospects than the liquid formulation for the control of larvae of vector mosquitoes.

KEY WORDS Thermal fogging, Bacillus thuringiensis israelensis, Aedes aegypti, Aedes albopictus, Anopheles dirus, Culex quinquefasciatus

INTRODUCTION

Control of vectorborne diseases, in particular dengue fever (DF) and dengue hemorrhagic fever (DHF), in Malaysia and other neighboring Southeast Asian countries follows World Health Organization (WHO) guidelines. House-to-house indoor thermal fogging sprays of diesel-based malathion (Cythion[®]; American Cyanimide, Princeton, NJ) are carried out when a DF or DHF outbreak occurs (Yap 1984; WHO 1995, 1997; Yap and Zairi 1999). Field evaluation of a few thermal fogging insecticide formulations against mosquitoes have been carried out in Malaysia (Yap et al. 1983, 1988, 1999, 2000, 2001) as well as in other tropical countries (Castle et al. 1999).

Microbial control of vector mosquitoes, such as by use of *Bacillus thuringiensis israelensis (Bti)*, is a relatively recent development. Several types of formulations are used in Malaysia (Foo and Yap 1982, 1983; Lee and Seleena 1992; Lee et al. 1996, 1997a). The effectiveness of *Bti* has been demonstrated in the control of mosquitoes such as *Culex* species (Foo and Yap 1982, Balaraman et al. 1983, Fry-O'Brien and Mulla 1996), *Aedes* species (Van Essen and Hembree 1980, Foo and Yap 1982, Klowden and Bullar 1984), *Anopheles* species (Foo and Yap 1982, Pantuwatana and Yougvanitsed 1984), and *Mansonia* species (Foo and Yap 1982, 1983).

The advantages of use of a microbial agent, in particular the bacterial agent Bti, for indoor vector control are that it can be used in drinking water, it

has adequate larvicidal activity, and it is nontoxic to nontarget organisms (WHO 1999).

Therefore, further development of such microbial agents is needed as an additional tool for the overall control program for vector mosquitoes. Recently, a new formulation of *Bti*, water-dispersible granules (WDGs; Vectobac ABG 6511), was evaluated as a larvicide against *Culex* species (Su and Mulla 1999). The objective of this study was to investigate the larvicidal efficacy of the WDG formulation granules in comparison with Vectobac 12AS[®] liquid, by using a portable thermal fogging sprayer in living premises on Penang Island, Malaysia.

MATERIALS AND METHODS

Mosquitoes: Laboratory-cultured, late 3rd- or early 4th-stage larvae of 4 mosquito species (Aedes aegypti (L.), Aedes albopictus (Skuse), Culex quinquefasciatus Say, and Anopheles dirus Peyton and Harrison) from the Vector Control Research Unit, Universiti Sains Malaysia, were used in this study.

Insecticide formulations and equipment: Samples of the WDG formulation (Vectobac ABG 6511, 3,000 international toxic unit [ITU]/mg) and liquid formulation (Vectobac 12AS, 1,200 ITU/mg) of *Bti* obtained from Abbott Laboratories (now Valent Biosciences Corporation, Libertyville, IL) were used for the tests. Two portable thermal foggers (Agrofog® model AF 35, Agro Technic Pte Ltd., Singapore) were used for larvicide applications. The discharge rate of the machines with nozzle size

per house for each bacterial formulat	tion in assessed pr	emises at Loror are the ra	ng Mahsuri, Bay tio of Vectobac	an Baru, Penan to seasoned wa	g Island, Malaysia ter.'	 The dilution rat 	es given in the	column heads
		Vectobac WD0	G (ABG 6511)			Vectobac	12AS®	
	1:20	1:40	1:80	1:160	1:8	1:16	1:32	1:64
Mean Vectobac formulation sprayed	968.6 g/ha	483.3 g/ha	237.5 g/ha	121.0 g/ha	2389.2 ml/ha	1215.5 ml/ha	597.3 ml/ha	303.9 ml/ha
Mean ITU per ha sprayed	2.91×10^{9}	1.45×10^{9}	0.71×10^{9}	0.36×10^{9}	$2.87 imes10^{9}$	$1.46 imes 10^{9}$	0.72×10^9	0.36×10^{9}
¹ WDG, water-dispersible granules; ITU, ir	nternational toxic uni	Ŀ						

Table 1. Summary of fogging time, discharge rate for Agrofog thermal fog machine (model AF35) with nozzle size 0.8, and mean Bacillus thuringiensis H-14 sprayed

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of 0.8 were set at 245 ± 5 ml/min. All spraying activities were conducted between 1800 and 1930 h.

Fogging operations and bioefficacy assessment: The trials were carried out in single-story terrace residential houses in an urban settlement situated in Bayan Baru, a town on the southwestern coastal area on Penang Island, Malaysia. The performance of the WDG formulation at dilution rates of 1:20, 1:40, 1:80, and 1:160 and the liquid formulation at dilution rates of 1:8, 1:16, 1:32, and 1:64 were compared. The assessments for each of the species were carried out with 20 larvae placed in a cylindrical paper cup (top diameter 8 cm and height 10 cm) filled with 200 ml of seasoned tap water at each checkpoint. Two checkpoints, 1 in the living room and the other in the kitchen, were set for each tested house. The test site consisted of more than 20 lanes of residential terrace houses. A minimum of 10 similar-sized single-story concrete houses (5 with assessment cups) at alternate positions (with 1 house in between) from a single lane were chosen for the spraying of each of the formulation. Different lanes, at least 50 m apart, were chosen for the spraying of each formulation. Each of the selected houses was sprayed for a period of 56.6 \pm 2.5 sec and an area of approximately 120 m² was covered. With the mean discharge rate of the machine at 245.0 ± 5.0 ml/min, the mean volume sprayed per house was 231.0 ± 3.0 ml.

The thermal fogging application of Bti formulations followed essentially the same technique used in the operational control of dengue vectors, namely, Ae. aegypti and Ae. albopictus, during a DF outbreak (WHO 1995, 1997). When conducting a house-to-house spray, a team of 2 persons is required. The 1st person operates the fogging machine and the 2nd person guides the 1st person and keeps track of spraying time. The team usually starts spraying from the back of the house (i.e., the kitchen area) and walks backwards until they reach the front of the house (i.e., the living room). The entire process of spraying from the back to the front of the house is done within a specified time. While spraying, the nozzle of the thermal fogging machine is pointed approximately 30° downwards and is swung constantly from side to side to ensure even coverage of the targeted house.

Larvae were brought back to the laboratory after 1 h of exposure and kept in clean paper cups. Mortality of larvae was recorded at 24 and 48 h after treatment. For comparative efficacy, the percentage of mortality values were subjected to an arcsine transformation followed by a comparison of means using the Duncan multiple range test (SAS Institute 1985). Temperature and percentage of relative humidity of indoor premises were recorded.

RESULTS AND DISCUSSION

Details of the sprays applied (dilutions, quantity, and ITUs applied) are shown in Table 1. The mean

Table 2. Mean percentalkitchen areas from 5 as	ge mortality ag sessed premise	gainst mosquito es situated at Lo Bacillus thuring	larvae of Aedes c rong Mahsuri, Ba tensis H-14 formu	<i>aegypti, Aedes al</i> ayan Baru, Penal ulations. No mor	bopictus, Culex ng Island, at 24 tality was recor	<i>quinquefasciatus</i> and 48 h after tr ded in the contro	t, and Anopheles eatment by thern I group. ¹	<i>dirus</i> in both liv nal fog spraying	ving rooms and with various
		Aedes	aegypti	Aedes a	lbopictus	Culex quin	quefasciatus	Anophel	es dirus
Formulations ²	Location	24 h	48 h	24 h	48 h	24 h	48 h	24 h	48 h
Vectobac WDG (ABG	Living room	$100.0 \pm 0.0^{\circ}$	100.0 ± 0.0^{a}	98.0 ± 1.0^{a}	$99.0 \pm 1.0^{\circ}$	100.0 ± 0.0^{a}	$100.0 \pm 0.0^{\circ}$	100.0 ± 0.0^{a}	100.0 ± 0.0^{a}
6511) diluted in 20-	Kitchen	100.0 ± 0.0^{a}	100.0 ± 0.0^{a}	100.0 ± 0.0^{a}	100.0 ± 0.0^{a}	100.0 ± 0.0^{a}	$100.0 \pm 0.0^{\circ}$	100.0 ± 0.0^{a}	100.0 ± 0.0^{a}
fold water (2.91 \times	Total	$100.0 \pm 0.0^{\circ}$	100.0 ± 0.0^{a}	$99.0 \pm 1.0^{\circ}$	99.5 ± 1.0^{a}	100.0 ± 0.0^{a}	100.0 ± 0.0^{a}	100.0 ± 0.0^{a}	100.0 ± 0.0^{a}
Vectohac WDG (ABG	Living room	$100.0 \pm 0.0^{\circ}$	100.0 ± 0.0^{a}	100.0 ± 0.0^{a}	100.0 ± 0.0^{a}	$98.0 \pm 1.0^{\circ}$	98.0 ± 1.0^{a}	100.0 ± 0.0^{a}	100.0 ± 0.0^{a}
6511) diluted in 40-	Kitchen	100.0 ± 0.0^{a}	100.0 ± 0.0^{a}	100.0 ± 0.0^{a}	$100.0 \pm 0.0^{\circ}$	$98.0 \pm 1.0^{\circ}$	99.0 ± 1.0^{a}	$82.0 \pm 6.4^{\circ}$	100.0 ± 0.0^{a}
fold water (1.45 \times 10° TTU/ha)	Total	100.0 ± 0.0^{a}	$100.0 \pm 0.0^{\mu}$	100.0 ± 0.0^{a}	100.0 ± 0.0^{a}	98.0 ± 1.0	98.5 ± 1.0^{a}	91.0 ± 3.2^{b}	100.0 ± 0.0^{a}
Vectohac WDG (ABG	Living room	52.0 ± 1.0^{d}	100.0 ± 0.0^{a}	$67.0 \pm 18.4^{\circ}$	98.0 ± 1.0^{a}	97.0 ± 1.0^{a}	100.0 ± 0.0^{a}	$84.0 \pm 4.8^{\rm b}$	97.0 ± 2.2^{a}
(511) diluted in 80-	Kitchen	50.0 ± 1.0^{d}	100.0 ± 0.0^{a}	$65.0 \pm 14.6^{\circ}$	98.0 ± 1.0^{a}	95.0 ± 1.0^{a}	$100.0 \pm 0.0^{\circ}$	82.0 ± 3.2^{b}	$95.0 \pm 1.4^{\circ}$
fold water $(0.71 \times 10^{9} \text{ trut})$	Total	51.0 ± 1.0^{d}	100.0 ± 0.0^{a}	$66.0 \pm 16.5^{\circ}$	$98.0 \pm 1.0^{*}$	$96.0 \pm 1.0^{\circ}$	100.0 ± 0.0^{a}	83.0 ± 4.0 ^b	96.0 ± 1.8^{a}
Vectobac WDG (ABG	Living room	$53.0 \pm 13.6^{\circ}$	$61.0 \pm 15.4^{\circ}$	$62.0 \pm 17.4^{\circ}$	$77.0 \pm 6.4^{\circ}$	$71.0 \pm 3.2^{\circ}$	87.0 ± 2.6^{b}	$94.0 \pm 2.2^{\circ}$	100.0 ± 0.0^{a}
6511) diluted in 160-	Kitchen	$70.0 \pm 14.2^{\circ}$	79.0 ± 13.6^{b}	47.0 ± 22.4^{d}	$61.0 \pm 12.4^{\circ}$	$70.0 \pm 4.6^{\circ}$	$77.0 \pm 4.6^{\rm b}$	$96.0 \pm 1.8^{\circ}$	98.0 ± 1.2^{a}
fold water (0.36 \times	Total	61.5 ± 13.9°	$70.0 \pm 14.5^{\circ}$	54.5 ± 19.9^{d}	69.0 ± 9.4°	$70.5 \pm 3.9^{\circ}$	82.0 ± 3.6^{b}	95.0 ± 2.0^{a}	99.0 ± 0.6^{a}
	T ining woom	1000 ± 0.03	1000 + 003	$000 + 10^{a}$	$00.0 + 1.0^{a}$	100.0 + 0.0a	$100.0 + 0.0^{\circ}$	100.0 + 0.0a	100.0 + 0.0a
Vectobac 12AS ^e diluted in 8-fold water	LIVING TOULI Kitchen	100.0 + 0.0	100.0 + 0.0	$100.0 \pm 0.0^{\circ}$	100.0 ± 0.0^{4}	$100.0 \pm 0.0^{\circ}$	$100.0 \pm 0.0^{\circ}$	100.0 ± 0.0^{a}	$100.0 \pm 0.0^{\circ}$
$(2.87 \times 10^9 \text{ FTU/ha})$	Total	$100.0 \pm 0.0^{\circ}$	100.0 ± 0.0^{a}	$99.5 \pm 0.5^{*}$	99.5 ± 0.5^{a}	100.0 ± 0.0^{a}	$100.0 \pm 0.0^{*}$	100.0 ± 0.0^{a}	100.0 ± 0.0^{a}
Vectobac 12AS diluted	Living room	93.0 ± 1.0^{a}	97.0 ± 2.2^{a}	94.0 ± 2.0^{a}	96.0 ± 2.0^{a}	$80.0 \pm 6.4^{\rm b}$	90.0 ± 3.6^{a}	96.0 ± 4.6^{a}	100.0 ± 0.0^{a}
in 16-fold water	Kitchen	92.0 ± 1.0^{a}	95.0 ± 3.2^{a}	96.0 ± 1.0^{a}	99.0 ± 1.0^{a}	82.0 ± 8.2^{b}	95.0 ± 5.4^{a}	89.0 ± 4.2^{b}	99.0 ± 1.0^{4}
$(1.46 \times 10^9 \text{ ITU/ha})$	Total	$92.5 \pm 1.0^{\circ}$	95.5 ± 2.7^{a}	95.0 ± 1.5^{a}	97.5 ± 1.5^{a}	81.0 ± 7.3^{b}	92.5 ± 4.5^{a}	$92.5 \pm 4.4^{\rm b}$	$99.5 \pm 0.5^{*}$
Vectobac 12AS diluted	Living room	$62.0 \pm 14.2^{\circ}$	$62.0 \pm 15.6^{\circ}$	$66.0 \pm 16.8^{\circ}$	76.0 ± 6.6^{b}	$65.0 \pm 12.4^{\circ}$	$77.0 \pm 5.5^{\circ}$	83.0 ± 8.4^{b}	99.0 ± 0.8^{a}
in 32-fold water	Kitchen	$79.0 \pm 13.2^{\circ}$	94.0 ± 5.2^{a}	83.0 ± 4.2^{b}	84.0 ± 4.2^{b}	$71.0 \pm 11.2^{\circ}$	$80.0 \pm 4.3^{\rm b}$	$86.0 \pm 6.6^{\circ}$	100.0 ± 0.0^{4}
$(0.72 \times 10^9 \text{ ITU/ha})$	Total	$70.5 \pm 13.7^{\circ}$	$78.0 \pm 10.4^{\circ}$	$75.0 \pm 10.5^{\circ}$	$80.0 \pm 5.4^{\circ}$	$68.0 \pm 11.8^{\circ}$	78.5 ± 4.9^{b}	84.5 ± 7.5^{b}	99.5 ± 0.4^{a}
Vectobac 12AS diluted	Living room	$60.0 \pm 13.4^{\circ}$	70.0 ± 13.8^{b}	$69.0 \pm 16.8^{\circ}$	71.0 ± 8.2^{b}	$34.0 \pm 22.6^{\circ}$	50.0 ± 12.2^{d}	55.0 ± 14.2^{d}	$68.0 \pm 12.6^{\circ}$
in 64-fold water	Kitchen	$59.0 \pm 25.2^{\circ}$	$62.0 \pm 24.2^{\circ}$	$73.0 \pm 14.6^{\circ}$	$83.0 \pm 4.2^{\circ}$	$74.0 \pm 16.2^{\circ}$	82.0 ± 6.4^{b}	62.0 ± 9.8^{d}	90.0 ± 1.2^{b}
$(0.36 \times 10^{9} \text{ ITU/ha})$	Total	$59.5 \pm 19.3^{\circ}$	$66.0 \pm 19.0^{\circ}$	$71.0 \pm 14.7^{\circ}$	77.0 ± 6.2^{b}	54.0 ± 19.4^{d}	$66.0 \pm 9.3^{\circ}$	59.5 ± 12.0^{d}	$79.0 \pm 6.9^{\circ}$
¹ Mean percentages mortali ² WDG, water-dispersible g	ty followed by th ranules; ITU, int	ne same letters wit ernational toxic ur	hin the same colum it.	uns are not significa	antly different (P	< 0.05, Duncan mu	ltiple range test).		

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temperature (27 \pm 1.2°C) and relative humidity (90 \pm 3.5%) that were recorded reflect the normal hot and humid indoor tropical environment. Vectobac ABG 6511 showed excellent larvicidal effects against all 4 mosquito species at 3 of the doses tested (2.91 \times 10°, 1.45 \times 10°, and 0.71 \times 10° ITU/ ha), with more than 96% mortality at 48 h after spraying. As a comparative formulation, Vectobac 12AS showed good larvicidal activity against all 4 mosquito species at 2 of the higher doses tested (2.87 \times 10° and 1.46 \times 10° ITU/ha), with more than 92.5% mortality at 48 h after spraying. However, Vectobac 12AS at 0.36 \times 10° ITU/ha was less effective against *Cx. quinquefasciatus* at 24 h after spraying (Table 2).

Larvae of An. dirus were significantly more susceptible than the other test species to both waterbased Vectobac ABG 6511 and Vectobac 12AS at lower doses $(0.36 \times 10^9 \text{ ITU/ha})$. Overall, results indicate that Vectobac ABG 6511 and Vectobac 12AS at higher doses $(1.45-2.91 \times 10^9 \text{ and } 1.46 2.87 \times 10^9$ ITU/ha) are highly effective larvicides against all 4 mosquito species (Table 2). Moreover, even at the lowest dose tested, Vectobac ABG 6511 $(0.36 \times 10^9 \text{ ITU/ha})$ and Vectobac 12AS $(0.36 \times$ 109 ITU/ha) showed some larvicidal properties, with more than 66% mortality at 48 h after spraying. The results in both of the spraying localities (kitchen and living room) were similar for both Vectobac formulations against all 4 mosquito species tested.

In conclusion, Vectobac ABG 6511 provided significantly better larvicidal activity when compared with Vectobac 12AS at similar doses (in terms of ITU/ha). This suggests that the new granule formulation is more effective as a larvicide against all 4 vector mosquito species tested and should be considered as an additional tool for vector control. Furthermore, the WDG formulation is a more suitable formulation for vector control in a tropical environment because of its easy shipment and longer shelflife when compared with liquid formulations.

In vector mosquito control programs, simultaneous control of adult and larval stages is preferable so as to reduce the overall vector mosquito population and subsequently reduce or disrupt disease transmission. In the case of dengue vector control, such an approach is essential because of transovarial transmission of dengue virus in *Ae. aegypti* and *Ae. albopictus* (Lee et al. 1997b).

In outdoor ultra-low-volume spray, tests recently have been carried out on the combination of chemical adulticides such as Pesguard[®] 102 (active ingredient [AI]: *d*-allethrin and *d*-phenothrin, both at 5% weight/weight [w/w]), Aqua Resigen[®] (AI: *s*bioallethrin 0.14%, permethrin 10.11%, and piperonyl butoxide 9.96% w/w/w), and malathion with *Bti* as larvicide so as to achieve both adult and larva control with 1 spray (Lee et al. 1997b; Yap et al. 1997a, 1997b). The possibility of achieving both larvicidal and adulticidal activity by integrating chemical and microbial agents with indoor thermal fogging spray should also be investigated.

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REFERENCES CITED

- Balaraman K, Balasubramaniam M, Manonmani LM. 1983. Bacillus thuringiensis H-14 (VCRC B-17) formulation as mosquito larvicide. Indian J Med Res 77: 33-37.
- Castle T, Amador M, Rawlins S, Figueroa JP, Reiter P. 1999. Absence of impact of aerial malathion treatment on *Aedes aegypti* during a dengue outbreak in Kingston, Jamaica. *Rev Panam Salud Publica Pan Am J Public Health* 5:100–105.
- Foo AES, Yap HH. 1982. Comparative bioassays of Bacillus thuringiensis H-14 formulations against four species of mosquitoes in Malaysia. Southeast Asian J Trop Med Public Health 13:1–5.
- Foo AES, Yap HH. 1983. Field trials on the use of Bacillus thuringiensis serotype H-14 against Mansonia mosquitoes in Malaysia. Mosq News 43:306-310.
- Fry-O'Brien LL, Mulla MS. 1996. Effect of tadpole shrimp, *Triops longicaudatus*, (Notostraca: Triopsidae), *Bacillus thuringiensis* var. israelensis in experimental microcosms. J Am Mosq Control Assoc 12:33–38.
- Klowden MJ, Bullar LA Jr. 1984. Oral toxicity of *Bacillus thuringiensis* subsp. *israelensis* to adult mosquitoes. *Appl Environ Microbiol* 48:665–667.
- Lee HL, Antonietta PE, Seleena P, Chiang YP. 1997a. Simultaneous ultra-low-volume application of adulticide (malathion) and larvicide (*Bacillus thuringiensis* H-14) for the control of dengue vectors. *Int Med Res J* 1:13– 19.
- Lee HL, Gregorio ER Jr, Khadri MS, Seleena P. 1996. Ultralow volume application of *Bacillus thuringiensis* ssp. *israelensis* for the control of mosquitoes. J Am Mosq Control Assoc 12:651–655.
- Lee HL, Mustafakamal I, Rohani A. 1997b. Does transovarial transmission of dengue virus occur in Malaysia Aedes aegypti and Aedes albopictus. Southeast Asian J Trop Med Public Health 28:230-232.
- Lee HL, Seleena P. 1992. Field larvicidal effect of a Malaysian isolate of *Bacillus thuringiensis* serotype H-14 against *Aedes albopictus* (Skuse). J Biosci 3:97-101.
- Pantuwatana S, Yougvanitsed A. 1984. Preliminary evaluation of *Bacillus thuringiensis* serotype H-14 and *Bacillus sphaericus* strain 1593 for toxicity against mosquito larvae in Thailand. J Sci Soc Thail 10:101–108.

- SAS Institute. 1985. SAS[®] user's guide: statistics, version 5 ed Cary, NC: SAS Institute, Inc.
- Su T, Mulla MS. 1999. Field evaluation of new waterdispersible granular formulations of *Bacillus thuringiensis* ssp. *israelensis* and *Bacillus sphaericus* against *Culex* mosquitoes in microcosms. J Am Mosq Control Assoc 15:356–365.
- Van Essen FW, Hembree SC. 1980. Laboratory bioassay of *Bacillus thuringiensis israelensis* against all instars of *Aedes aegypti* and *Aedes taeniorhynchus* larvae. *Mosg News* 40:424–431.
- WHO [World Health Organization]. 1995. Guidelines for dengue surveillance and mosquito control. WHO/ WPRO. Geneva, Switzerland: World Health Organization.
- WHO [World Health Organization]. 1997. Dengue haemorrhagic fever, diagnosis, treatment, prevention and control 2nd ed. Geneva, Switzerland: World Health Organization.
- WHO [World Health Organization]. 1999. *Bacillus thuringiensis* Environmental health criteria 217. Geneva, Switzerland: World Health Organization.
- Yap HH. 1984. Vector control in Malaysia—present status and future prospects. *J Malays Soc Health* 4:7–12.
- Yap HH, Chong ASC, Adanan CR, Chong NL, Rohaizat B, Abdul Malik Y, Lim SY. 1997a. Performance of ULV formulations (Pesguard® 102/Vectobac® AS 12) against three mosquito species. J Am Mosq Control Assoc 13:384–388.
- Yap HH, Chong ASC, Chong NL, Adanan CR, Abdul Malik Y, Lim SY, Rohaizat B. 1997b. ULV field studies of combined spraying of Aqua Resigen[®] and Vectobac[®] AS12 against Aedes aegypti, Aedes albopictus and Culex quinquefasciatus. Trop Biomed 14:57-63.

- Yap HH, Khoo TC, Chee CS, Cheong WH. 1983. Comparative adulticidal and larvicidal effects of thermal fogging formulations of Dursban, malathion, and bioresmethrin against Aedes aegypti (Linnaeus) and Culex quinquefasciatus (Say) on Penang Island, Malaysia. In: Pang T, Pathmanathan, eds. Dengue/dengue haemorrhagic fever Proceeding of the International Conference. 1983 September 1–3; University of Malaya, Kuala Lumpur, Malaysia. p 231–246.
- Yap HH, Khoo TC, Tan HT, Chung KK, Abdul Malik Y, Narayanan VS. 1988. Comparative adulticidal and larvicidal effects of thermal fogging formulations of Resigen[®] and malathion against *Aedes aegypti* (Linnaeus) and *Culex quinquefasciatus* Say in urban areas, Malaysia. *Trop Biomed* 5:125–130.
- Yap HH, Lee YW, Chong ASC, Zairi J. 1999. Bioefficacy of pyrethroid formulation (Pesguard[®] PS 102) in thermal fogging application against urban mosquito species of public health importance. *Trop Biomed* 16:17–22.
- Yap HH, Lee YW, Zairi J. 2000. Chemical control of mosquitoes. In: Ng FSP, Yong HS, eds. Mosquitoes and mosquito-borne diseases: biology, surveillance, control, personal and public health measures Kuala Lumpur, Malaysia: Academy of Sciences Malaysia. p 197–210.
- Yap HH, Lee YW, Zairi J, Jahangir K, Adanan CR. 2001. Indoor thermal fogging application of Pesguard® FG 161, a mixture of *d*-tetramethrin and cyphenothrin, using portable sprayer against vector mosquitoes in the tropical environment. J Am Mosq Control Assoc 17:28– 32.
- Yap HH, Zairi J. 1999. Mosquito control. In: Lee CY, Yap HH, Chong NL, Zairi J, eds. Urban pest control, a Malaysian perspective Penang, Malaysia: Universiti Sains Malaysia. p 39–49.