

small numbers along the edges of the main canals, and in large numbers in the concrete drop structures (fig. 1). A total of 206 drop structures was located and examined in the survey area, of which 193, or 93 per cent, contained Simuliidae. Collections of *S. vittatum* and *S. griseum* larvae and pupae were made from several of the drops on July 22 and August 14, 1953.

In this particular area, irrigation water is delivered to the user under the rotation method whereby he receives water for definite periods of time and is without water during the intermediate periods. Consequently water was sometimes shut out of laterals for a period of several days. When this happened sufficient water usually remained in the concrete apron of each drop structure to permit the blackfly larvae and pupae to survive until the water was turned into the lateral again.

When water was not flowing in the lateral it was possible to estimate the density of the Simuliidae population in the concrete aprons. This was done by measuring a 1-foot square area on the side of an apron and counting the number of larvae and pupae attached to the concrete. In late August two 1-foot square areas were sampled in 10 different aprons.

The larvae and pupae in these samples ranged in number from 2 to 37 with an average of 18 per square foot. Each apron was found to measure approximately 28.5 square feet so that the breeding potential of a single drop structure was 513 Simuliidae. On this basis it is estimated that the 206 drops contained in excess of 105,000 blackflies.

The blackflies were identified by Dr. Alan Stone of the Division of Insect Detection and Identification, Bureau of Entomology and Plant Quarantine, Washington 25, D. C.

A TECHNIQUE TO FACILITATE IDENTIFICATION OF MOSQUITOES IN LIGHT TRAP COLLECTIONS

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Light traps are used extensively for sampling adult mosquito populations; however, the large catches which are frequently taken make the routine task of identifying and counting the specimens a time-consuming and tedious process. The technique described below has greatly facilitated the processing of light trap collections of mosquitoes which have been made in connection with studies undertaken in irrigated areas of several western states.

The technique involves the use of (1) a wide-field stereoscopic microscope, (2) a manually operated stage consisting of a

base plate and sliding horizontal trough and (3) a nine-unit laboratory counter. The base plate of the manual stage is constructed so that it replaces the removable glass stage on the microscope. Mosquitoes from the light trap collections are thinly scattered over the bottom of the trough which operates in the center of the field. By slowly moving the trough beneath the objective the mosquitoes may be identified and recorded on the laboratory counter.

¹ #A-2030 Clay-Adams Co., Inc. The trade name is used as a means of identifying the product. Its use does not constitute endorsement by the U. S. Public Health Service.

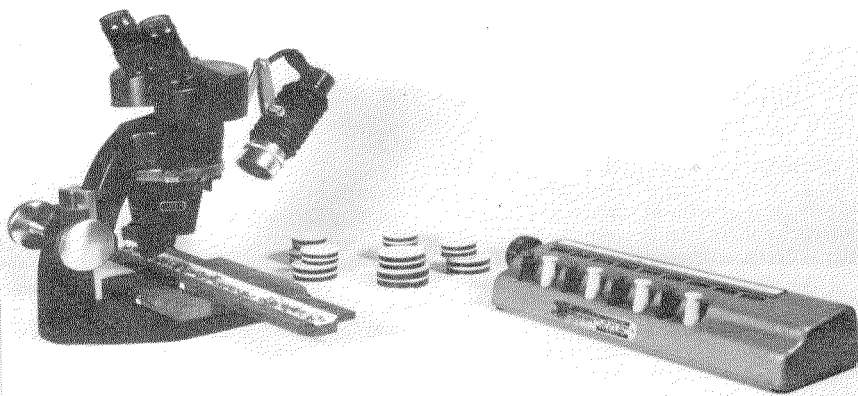


FIG. 1. Stereoscopic microscope equipped with manual stage and nine-unit laboratory counter used for identifying and counting mosquitoes from light-trap collections.

fig. 1). Males and females of four species and their total can be recorded on the nine-unit counter. With a little practice, the counter can be operated by touch, allowing undivided attention to be devoted to the microscope field.

Construction details of the microscope stage are shown in figure 2. The trough and base plate are constructed of $\frac{1}{8}$ " and $\frac{1}{4}$ " tempered masonite glued together. The trough used in this laboratory is 12 inches long with 1-inch divisions. These divisions are useful in determining which mosquitoes have been identified in the event of an interruption. The width of the trough is dependent on the field covered by the combination of objectives and eye-pieces normally used for mosquito identification. The sides of the trough are sloped to prevent shadows and the bottom may be painted white or black, depending on individual preference.

It has been estimated that the time required to identify and count the mos-

quitoes in the larger trap collections is reduced as much as 50 per cent by the use of this technique. The technique removes much of the tedium of mosquito counting and reduces the handling of individual specimens. It may be used for unsorted trap collections where sorting, identifying, and counting are all accomplished in one operation. This technique is adaptable for use in handling other kinds of insects collected in light trap samples.

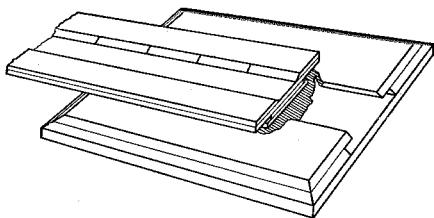


FIG. 2. Perspective view showing construction details of the manually operated microscope stage.