

FIELD OBSERVATIONS ON THE NECTAR FEEDING HABITS OF SOME MINNESOTA MOSQUITOES¹

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INTRODUCTION. The literature contains reports of a number of casual daylight observations of mosquito visitation to flowers. Robertson (1889, 1928) and Knab (1907) were among the earliest to report such activities. Raup (1930), Britten (1937), Philip (1943), Larsen (1948), and Nielson and Greve (1950) recorded similar findings, but from these early records it is often difficult to determine whether an actual nectar meal was taken. West and Jenkins (1951), through the use of radioactive phosphorus, proved in the laboratory that *Aedes communis* (DeGeer) did take up nectar from flowers. Hocking (1953) thinks floral nectar furnishes most of the energy used in flight. More thorough field studies have been made recently by Haeger (1955) on a salt marsh mosquito and by Breeland and Pickard (1961) on mosquito visits to the flower heads of *Eupatorium* and *Solidago*. There has been, however, no extensive published study on the nocturnal nectar feeding habits of mosquitoes in nature. It is our purpose here to present such observations made throughout the summer in Minnesota.

METHODS AND OBSERVATIONS. Mosquitoes were observed and collected from twilight up to midnight from early June through September, 1962. During this period 102 collections were made, each representing the capture of one or more mosquitoes taking nectar from a plant species for one evening. A flashlight emitting red light was used to detect and observe the mosquito activity; the mosquitoes were seemingly insensitive to the red light as it did not disturb their activity in

any noticeable manner. Once a mosquito had been observed to be taking nectar, an aspirator was used to remove it from the flower of the plant; all resting mosquitoes were ignored. After capture, the mosquitoes were transferred from the aspirator to a killing jar and retained for identification in the laboratory. The majority of the observations were made at the University of Minnesota Arboretum located about 20 miles west of Minneapolis, since this area afforded access to a large variety of flowering plants. Supplementary records were made in the northern and southern central portions of the state.

MOSQUITOES AND THE PLANTS FROM WHICH NECTAR MEALS WERE TAKEN (numbers represent males and females, respectively) *Aedes vexans* (MEGALOPTERUS): (825, 702), *Apocynum androsaemifolium* (1, 6), *Aquilegia chrysantha* (0, 1), *Asclepias syriaca* (149, 155), *Aster alpinus* (1, 0), *Catalpa bignonioides* (2, 5), *Cirsium arvense* (15, 18), *C. lanceolatum* (2, 4), *Cornus stolonifera* (0, 5), *Echinocystis lobata* (4, 36), *Eupatorium purpureum* (8, 8), *Heracleum lanatum* (2, 1), *Hydrangea arborescens inscolor* (8), *H. radiata* (2, 4), *Lilium* "emerald trumpet" (19, 17), *L.* "regal" (16, 19), *L. superbum* (3, 0), *Melilotus alba* (40), *Nepeta cataria* (12, 8), *Pastinaca sativa* (2, 2), *Philadelphus californicus* (9), *P. caucasicus* (6, 15), *P. coronarius*, *Phlox paniculata* (45, 24), *Physocarpus opulifolius* (0, 2), *Potentilla fruticosa* (2, 8), *Rosa* "Sir Thomas Lipton" (0, 14), *Rudbeckia subtomentosa* (0, 6), *Solidago canadensis* (31, 72), *Sonchus oleraceus* (6), *Spiraea bumalda* (12, 11), *S. colloidalis* (15, 10), *S. latifolia* (335, 81), *S. rosae* (14, 7), *S. salicifolia* (0, 5), *Symphoricarpos occidentalis* (3, 4), *Syringa amurensis japonica* (19, 39), *Tamarix pe-*

¹ Paper No. 4961, Scientific Journal Series, Minnesota Agricultural Experiment Station, St. Paul 1, Minnesota. This study was supported in part by a summer fellowship from the National Science Foundation.

dra (16, 14), *Trifolium pratense* (20, 1), *T. repens* (16, 13).

AEDES CINEREUS (MEIGEN): (9, 1), *Asclepias syriaca* (1, 2), *Cirsium arvense* (0, 3), *Cornus stolonifera* (0, 3), *Chinocystis lobata* (0, 1), *Lilium "regal"* (3), *Philadelphus californicus* (2, 11), *caucasicus* (3, 6), *Rosa "Sir Thomas Mott"* (0, 1), *Solidago canadensis* (0, 6), *Spiraea latifolia* (1, 0), *S. salicifolia* (0, 1), *Thymophoricarpus occidentalis* (2, 0), *Syngga amurensis japonica* (0, 2), *Verbena stricta* (0, 1).

AEDES FITCHII (FELT AND WILSON): (1, 16), *Chrysanthemum* sp. (1), *Fraxinus pennsylvanica lanceolata* (1), *Philadelphus caucasicus* (0, 1), *coronarius aureus* (0, 1), *Potentilla fruticosa* (0, 1), *Rosa "Sir Thomas Mott"* (0, 5), *Spiraea latifolia* (0, 1), *Syngga amurensis japonica* (0, 4), *Trifolium ens* (0, 1).

AEDES TRIVITTATUS (COQUILLET): (7, 4), *Apocynum androsaemum* (0, 2), *Catalpa bignonioides* (3, 0), *gnus stolonifera* (1, 0), *Phlox paniculata* (0), *Tamarix pentandra* (0, 2).

AEDES PUNCTOR (KIRBY): (0, 8), *Heracleum lanatum* (0, 1), *Solidago canadensis* (0, 3), *Spiraea salicifolia* (0, 1), *Verbena stricta* (0, 3).

AEDES STICTICUS (MEIGEN): (0, 1), *Heracleum lanatum* (0, 1), *Spiraea salicifolia* (0, 2), *Verbena stricta* (0, 2).

AEDES CANADENSIS (THEOBALD): (0, 3), *Chrysanthemum* sp. (0, 1), *Heracleum lanatum* (0, 1), *Solidago canadensis* (0, 1).

AEDES DORSALIS (MEIGEN): (0, 1), *Asclepias syriaca* (0, 2), *Phlox paniculata* (0, 1).

AEDES ABSERRATUS (FELT AND WILSON): (0, 2), *Physocarpus opulifolius* (1), *Trifolium hybridum* (0, 1).

AEDES EXCRUCIANS (WALKER): (1), *Heracleum lanatum* (0, 1).

MANSONIA PERTURBANS (WALKER): (165, 135), *Apocynum androsaemifolium* (0, 1), *Heracleum lanatum* (0, 2), *Lilium "emerald trumpet"* (1), *L. "regal"* (0, 3), *Nepeta cataria* (5), *Philadelphus californicus* (9, 3),

P. caucasicus (3, 2), *P. coronarius aureus* (12, 6), *Phlox paniculata* (0, 1), *Potentilla fruticosa* (0, 12), *Spiraea latifolia* (18, 42), *Syringa amurensis japonica* (123, 57).

CULEX RESTUANS (THEOBALD): (12, 10), *Asclepias syriaca* (2, 0), *Catalpa bignonioides* (2, 0), *Chrysanthemum* sp. (1, 0), *Eupatorium purpurum* (0, 2), *Melilotus alba* (0, 2), *Philadelphus caucasicus* (1, 0), *Solidago canadensis* (0, 3), *Spiraea latifolia* (6, 3).

CULEX TARSALIS (COQUILLET): (0, 2), *Solidago canadensis* (0, 2).

CULISETA INORNATA (WILLISTON): (12, 19), *Heracleum lanatum* (0, 1), *Melilotus alba* (0, 3), *Philadelphus caucasicus* (0, 1), *Physocarpus opulifolius* (0, 1), *Potentilla fruticosa* (1, 0), *Solidago canadensis* (0, 6), *Spiraea latifolia* (2, 3), *Syringa amurensis japonica* (9, 4).

There were 1,981 specimens distributed among 10 species of *Aedes*, two of *Culex*, and one each of *Culiseta* and *Mansonia*, in many instances with both sexes participating in the feeding. *Ae. vexans* provided the greatest numbers of mosquitoes (1,527) and the widest range of plant species (39) involved; this is as would be expected, since this mosquito is the most consistently abundant species over much of the state. Of special interest is the apparent wide choice of plants from which a nectar meal may be obtained and the ease with which mosquitoes may be found and observed in the act of imbibing nectar.

The feeding activity itself is worthy of comment. In most cases a probing action preceded the actual feeding. Although the feeding time as such is not very long, if a flower cluster is present the mosquito may continue to probe and feed over the entire flower area. As the nectar is being imbibed, the abdomen may be observed to be pulsating and swelling gradually. On two occasions nectar-fed females were taken that still had the remains of a partially digested blood meal. Once satiated, the mosquito will rest and engage in much rubbing of its hind legs. Several photographs of mosquitoes feeding on flowers at night are shown in Figures 1-3.

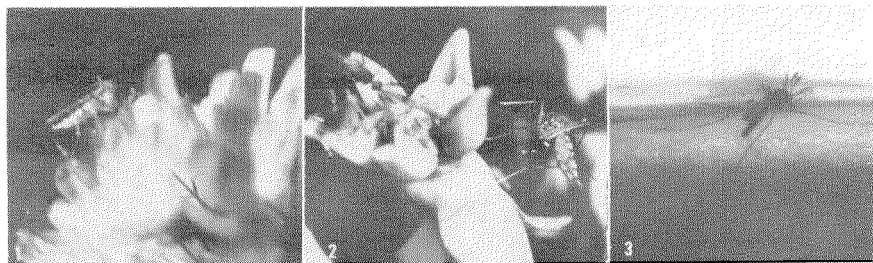


FIG. 1 (Left).—*Aedes vexans* female feeding on head of *Trifolium pratense*. FIG. 2 (Center).—*Aedes vexans* females on *Asclepias syriaca*. FIG. 3 (Right).—*Ae. vexans* male feeding inside the flower of regal lily.

Very few mosquitoes could be found feeding before dark. On nights when swarming was observed, heavy feeding on flowers invariably followed. For instance, on July 17, after swarming had taken place at a site near several *Spiraea latifolia* bushes, 20 male and female mosquitoes were collected from a single flower cluster during one 15-minute period and 41 during the following 15 minutes. The flowers were literally covered with mosquitoes.

Another example of such heavy feeding occurred on June 25 on the Japanese lilac, *Syringa amurensis japonica*. From 9:00–9:15 (CDT), only one *Ae. vexans* was taken; from 9:16–9:31, five mosquitoes representing this species as well as *Ae. fitchii* and *Mansonia perturbans* were captured. Then feeding increased dramatically, with 24 specimens, including the above 3 species plus *Culiseta inornata*, captured from 9:33–9:48. Thirty-three specimens of these four species were taken in only a 5-minute period between 10:00–10:15, as feeding reached its climax.

The first hour or so after dark would consistently produce the greatest numbers feeding at any one time, but if conditions were favorable, feeding did not completely stop during the entire observation period. Contrasted to the above observations, Downes (1958) reported that three northern *Aedes* species showed males predominating in the afternoon and females late in the evening.

Ae. cinereus was one of the earliest mos-

quitoes to feed. This species was often collected before it had become completely dark. Once it was dark, any of the other species might be collected. As one would anticipate, too much wind or temperature down in the 50° F. range reduced the feeding activity; also rainfall, even if it had occurred two or three hours before dark, seemed to detract from feeding. The temperature effect was particularly evident on many cool September nights; when temperatures dropped into the forties no collections could be made.

One of the most interesting observations concerned the greater numbers of mosquitoes feeding on the lighter colored flowers. The white, pink and yellow flowers standing side by side with darker ones produced feeding mosquitoes while the darker ones proved negative or actually so. For example, red clover (*Trifolium pratense*) would be negative while the white clovers (*Trifolium repens* and *Melilotus alba*) would produce numbers of feeding mosquitoes. Only when the moon was in its full phase and the sky was clear did mosquitoes appear in any numbers on such plants as red clover and phlox.

The availability of the nectar in a flower must also be an important factor for feeding; yarrow, some of the multi-petaled roses, and similar flowers proved negative even though the flowers are light colored. It is uncertain how important flower aroma may be to the attraction

mosquitoes. They do feed very heavily on the fragrant blossom of the common milkweed (*Asclepias syriaca*); 25 *Ac. vexans* were collected from one flower cluster of milkweed in 15 minutes. The age of the flower was also a factor, with fewer feeding attempts being observed on the older blossoms.

SUMMARY AND CONCLUSIONS. A total of 1,981 specimens representing 4 genera and 14 species of mosquitoes was captured in nature after imbibing nectar meals. They were taken from a large number of different plant species during the early light-time hours. Various factors, such as temperature, wind and rainfall, in addition to availability of the nectar, and the color, age and aroma of the flower, may affect the plant-feeding activity of the mosquito. The ease with which nectar-feeding mosquitoes are found and the variety of plants upon which they feed supports the belief that mosquitoes in nature may rely extensively upon nectar for satisfying their biological requirements.

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THE INFLUENCE OF FLOWER SCENTS ON AGGREGATIONS OF CAGED ADULT *AEDES AEGYPTI*

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INTRODUCTION. Female mosquitoes have been observed to visit and probe several species of flowering plants for nectar and nectars (Barr, 1958). Under laboratory conditions *Aedes communis* (De Geer) males and females visit flowers and ingest plant nectars and nectars as shown by their accumulation of radioactivity from flowering plants treated with P^{32} (West and Jenkins, 1951).

This behaviour is undoubtedly a factor in longevity of adult mosquitoes. In our preliminary experiments we observed that *Aedes aegypti* (Linnaeus) adults survived only 24-48 hours without water and 4 to 5 days on water without sugar whereas they can survive about 60 days in the laboratory when provided with sugar syrup or honey. Hocking (1953) has shown that nectar supply is important for