

g range evaluation of the dispersal pattern of these two salt marsh species.

SUMMARY. From the release of approximately 3 million P³²-tagged *Aedes taeniorhynchus* near Savannah, Georgia, 415 males and 29 females were recovered in 20 traps located at distances up to 20 miles from the departure point. Recoveries beyond 4 miles did not occur until the fourth night after release. Female specimens were captured at distances up to 18 miles but more than 75 percent of recoveries, exclusive of one trap 0.1 mile from the release point, were within

4 miles. The maximum distance at which males were trapped was 2 miles from the departure point.

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FIELD TESTS ON THE PERSISTENCE OF MOSQUITO LARVICIDES IN ALKALINE WATER IN CALIFORNIA

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INTRODUCTION. The frequency of application of mosquito larvicides in control programs in certain areas of California depends in part on the persistence of the insecticide in waters which are highly alkaline. When chlorinated hydrocarbon insecticides were used prior to the development of resistance, one larvicidal application often remained effective for several days (Robinson, 1946). Miles (1960) reported on the residual effectiveness of several chlorinated hydrocarbons for 2 to 3 seasons against floodwater mosquitoes.

Residue problems on agricultural crops and the development of physiological resistance have precluded the use of chlorinated hydrocarbons for mosquito larviciding in most California abatement districts; as a consequence organophosphorus compounds have become widely used in mosquito control programs.

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Organophosphorus compounds vary considerably in the rate with which they are hydrolyzed by alkaline water. Easily hydrolyzed insecticides frequently are not effective in the field even at high dosage rates, and under certain conditions they remain toxic to mosquitoes for only a few hours. To plan the most effective frequency of applications, the stability of organophosphorus compounds and other mosquito larvicides must be evaluated under field conditions.

METHODS AND MATERIALS. Since irrigated pastures were not suitable for field studies, a pilot study area was provided at Traver with the cooperation of the United States Department of Agriculture, the Delta Mosquito Abatement District, and the Kaweah Delta Gun Club of Traver.

In these 10' x 136' (1/32 acre) test plots, variables such as depth of water and movement of water from one plot to another were controlled. A natural cover of

TABLE 1.—24 hr. percent mortality of 4th instar *Culex tarsalis* after specific time interval.

Compound	Application (lb./acre)	Time after application for placement of larvae					
		Immed. aft.	4 hrs.	8 hrs.	24 hrs.	48 hrs.	72 hr
DDT E.C.*	1.0	100	100	100	71
Malathion E.C.	0.5	100	80	45
Malathion bentonite 20/30 mesh granules	0.5	72	37
Hercules 7522 E.C.	0.75	99	5
Dimethrin	0.3	88	43	32
Parathion E.C.	0.1	100	100	100	80	2	..
Parathion 30 mesh sand core granules	0.1	100	100	100	100	3	..
Baytex® E.C.	0.1	100	100	100	75
Baytex® 30 mesh sand core granules	0.1	100	100	100	55
Cynem® E.C.	0.1	100	100	100	80	61	..
Methyl Trithion® E.C.	0.25	100	100	100	70	36	..
Trithion® E.C.	0.5	100	100	100	100	100	99

* Emulsion concentrate.

grasses, weeds, and sedges developed on the sides and bottom of each plot. The plots were filled with water to a depth of about six inches before the application of insecticides and records of water temperature and pH were kept.

Emulsion concentrate sprays were diluted with one-half gallon of water and applied with a one-gallon B and G compression sprayer fitted with a 3004 Teejet nozzle on the wand. Granules were applied with a horn seeder (Raley, 1961).

Immediately after treatment of each plot, fourth instar *Culex tarsalis* Coquillett larvae were placed within the confines of a floating basket (tea strainer). An untreated plot with larvae in floating baskets served as a control. Negligible mortality was observed in the control baskets. Four baskets with 25 larvae in each were placed at random in each plot. At various intervals after treatment, fresh larvae were placed in other baskets set into the plots to determine if the water was still toxic. Mortality counts were made after 24 hours on all time sequence tests. Results of the tests are presented in Table 1. Each figure represents the average of three replications. The chemical compositions of the tested compounds are given in Table 2.

RESULTS. The study extended from March into the first week of June, 1961.

During this period the water temperature varied little; the mean temperature was 50° F. The average pH was 8.8. Drastic changes in temperature and pH may affect the residual activity of insecticides, noted by Mulla and Axelrod (1960).

Under the test conditions DDT, parathion (both granular and emulsion concentrate), Hercules 7522, and dimethrin were rapidly hydrolyzed. Trithion® emulsion concentrate at 0.5 lb/acre was

TABLE 2.—Chemical composition of tested compounds.

DDT	1,1,1 - trichloro - 2,2 - (<i>p</i> -chlorophenyl) ethan
Parathion	O,O-diethyl O- <i>p</i> -nitrophenyl phosphorothioate
Malathion	S - [1,2 - bis(ethoxyethyl)ethyl]O,O - methyl phosphorodithioate
Hercules 7522	2-chloro-5-isopropylphenyl N-methylcarbamate
Baytex®	O,O-dimethyl O-[4(methylthio)- <i>m</i> -tolyl] phosphorothioate
Methyl Trithion®	O,O-dimethyl S- <i>p</i> -chlorophenylthiomethyl phosphorodithioate
Trithion®	S[(<i>p</i> - chlorophenylthiomethyl)O,O-diethyl phosphorodithioate
Cynem®	O,O-diethyl O-(2-pyrazinyl)phosphorothioate
Dimethrin	2,4-dimethylbenzyl chrysothemumate

most persistent material tested. Parathion 30-mesh sand core granules persisted slightly longer than the emulsion concentrate applied at the same rate. On the other hand, Baytex® emulsion concentrate persisted slightly longer than 30-mesh sand core granules applied at the same rate.

These results indicate that both the formulation of insecticide and the chemical nature of the compound affect residual activity. Furthermore, the rate of hydrolysis in the field may be an important factor in persistence.

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MOSQUITOES AND OTHER ARTHROPODS FOUND IN BAGGAGE COMPARTMENTS OF INTERNATIONAL AIRCRAFT

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INTRODUCTION. The potential hazard of aircraft introducing an exotic arthropod or arthropod-borne disease into the United States has long been recognized. Routine arthropod surveillance of airports and appropriate control measures on aircraft entering the United States from foreign countries are applied by the entomological staff of the Division of Foreign Quarantine, U. S. Public Health Service.

While there have been numerous reports in the literature (Welch, 1939; Whitfield, 1939; Dethier, 1946; Denning *et al.*, 1947; Miller *et al.*, 1947; Hughes, 1949; Baird, 1951; Hughes *et al.*, 1956; and Porter, 1958) of arthropods recovered from aircraft, little has been published with reference to the comparative significance of the various compartments. The purpose of this study made in 1960 and 1961 was to evaluate the importance of the baggage compartments as a carrier of

arthropods, especially mosquitoes, in international traffic entering New Orleans, Louisiana; Honolulu, Hawaii; and Miami, Florida.

Griffitts and Griffitts (1931) found that the front baggage compartments of the Sikorsky amphibian aircraft were ideal for harboring mosquitoes. The majority of mosquitoes recovered by these authors were from this part of the airplane. In inspections of aircraft at Khartoum, Sudan, from 1935 to 1938, Whitfield (1939) found most of the insects in the main cabin, but reported that occasionally specimens were collected from the baggage compartments.

METHODS. The baggage compartments and cabins of 210 aircraft were inspected for arthropods at the Moisant International Airport in New Orleans. These planes were all four-motored propeller-type aircraft, each with two baggage compartments.

A total of 89 aircraft was inspected at the Honolulu International Airport. All of these planes were four-motored jet-type aircraft, each with two baggage compartments.

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