ends of the autosomes. III—L is very close to their drawing and III—R compares well along most of the length. It is more difficult to compare our maps of II with their drawings.

It is of interest that the two inversions described by Frizzi and DeCarli agree almost exactly in location with ours. We are convinced that we have the same inversions as described by them. Their stock of *freeborni* was obtained from Rozeboom; this is in all probability the same stock which we have (Ward and Kitzmiller, 1963). One difference seems to be in the frequency of the inversion in III—L. We have found this only once; evidently it was quite frequent in their stock.

In addition to the homologies of certain areas between *freeborni* and *atroparvus* mentioned above, there are certain clear homologies with other species. III—L is, as far as we can tell, identical or very similar in *atroparvus, freeborni,* and *punctipennis.* III—L in *earlei, crucians, aztecus,* and *occidentalis* are also very similar to *freeborni.* Preliminary observations clearly indicate the similarities.

The mitotic and meiotic chromosomes of *A. freeborni* consist of one pair of sub-telocentric heterosomes, and two pairs of metacentric autosomes (Fig. 2A). One pair of autosomes has arms of approximately equal length; we consider this to correspond with salivary chromosome II which also has approximately equal arms. The other pair of autosomes clearly has one arm shorter than the other; this corresponds well with salivary chromosome III. The X-chromosome in the mitotic configuration is as long as either autosome. Yet in the salivary preparations, the X is very much shorter, only about one-quarter the length of the autosomes. It is possible that the salivary X corresponds only to the shorter arm of the mitotic X, and that the longer portion, presumably heterochromatic, of the mitotic X, does not appear in salivary preparations.

SUMMARY. Salivary chromosome maps are presented for *Anopheles freeborni.* All chromosomes are mapped as homozygotes, and it is proposed that this be used as the "standard" map for this species. Supported in part by grant E 3486 USPHS.

Literature Cited

FRENCH, W. L., BAKER, R. H., and KITZMILLER, J. B. 1962. Preparation of mosquito chromosomes. Mosq. News 22:377–383.

FRIZZI, G. 1947. Cromosomi salivari in *Anopheles maculipennis.* Sci. Gen. 3:69–79.

FRIZZI, G., and DECARLI, L. 1954. Studio preliminare comparativo genetico e citogenetico fra alcune specie nordamericane di *Anopheles maculipennis* e l'*Anopheles maculipennis atroparvus Italiano.* Symposia *genetica* II:184–206.

KITZMILLER, J. B. 1953. Mosquito genetics and cytogenetics. Rev. Bras. Mal. Doen. Trop. 5:285–359.

———. 1963. Mosquito Cytogenetics. Bull. W.H.O. (in press).

WARD, R. A., and KITZMILLER, J. B. 1963. A list of laboratory colonies of anopheline mosquitoes. Mosq. News (in press).

CORRECTION

In the paper entitled "The Anatomy of a Naturally Occurring Sterile Adult Female *Aedes aegypti* (Linnaeus)" (Vol. 23, No. 2, June, 1963, page 165) by Jack Colvard Jones, line 16 in Column 1 continues in Column 2, lines 1 through 11. Also, line 24 in Column 1 continues in Column 2, lines 12 through 26. The author would also like to add that Spielman (Spielman, A., 1957: "The inheritance of autogeny in the *Culex pipiens* complex of mosquitoes." Amer. J. Hyg. 65:404–425) was apparently the first to report cases of ovarian atrophy, "apparently without developed follicles," in a mosquito *(Culex pipiens).* He found 51 cases of bilateral ovarian atrophy out of 6,000 females (see his paper, page 409).—J. C. J.