

- fornia group arboviruses: Immunodiffusion studies. *J. Immunol.* 99:276-284.
- Pantuwatana, S., W. H. Thompson, D. M. Watts, T. M. Yuill and R. P. Hanson. 1974. Isolation of LaCrosse virus from field collected *Aedes triseriatus* larvae. *Am. J. Trop. Med. Hyg.* 23(2):246-250.
- Sever, J. L. 1962. Application of a microtechnique to viral serological investigations. *J. Immunol.* 88:320-329.
- Sudia, W. D. and R. W. Chamberlain. 1967. Collection and processing of medically important arthropods for arbovirus isolation. Public Health Service, Center for Disease Control, Atlanta, Georgia. 29 pp.
- Sudia, W. D., V. F. Newhouse, C. H. Calisher and R. W. Chamberlain. 1971. California group arboviruses: Isolations from mosquitoes in North America. *Mosq. News* 31(4):576-600.
- U. S. Department of Health, Education, and Welfare, Public Health Service. 1962. Diagnostic complement fixation method (LBCF), CDC Laboratory Branch Training Manual.
- Watts, D. M., C. D. Morris, R. E. Wright, G. R. DeFoliart and R. P. Hanson. 1972. Transmission of LaCrosse virus (California encephalitis group) by the mosquito *Aedes triseriatus*. *J. Med. Entomol.* 9(2):125-127.
- Watts, D. M., S. Pantuwatana, G. R. DeFoliart, T. M. Yuill and W. H. Thompson. 1973. Transovarial transmission of LaCrosse virus (California encephalitis group) in the mosquito, *Aedes triseriatus*. *Science* 182(4117):1140-1141.

EXPERIMENTAL HYBRIDIZATION BETWEEN *Aedes sollicitans* (WALKER) AND *Aedes mitchellae* (DYAR)

GEORGE F. O'MEARA, JUDY W. KNIGHT AND DAVID G. EVANS

Florida Medical Entomology Laboratory, Florida Division of Health,
P.O. Box 520, Vero Beach, Florida 32960

ABSTRACT. *Aedes sollicitans* and *Aedes mitchellae* were hybridized using induced mating techniques. Viable, fertile, F₁ hybrids were obtained from reciprocal crosses between the two species. These F₁ hybrids were successfully backcrossed to both parental species. In general, each backcross progeny had a normal sex ratio with minimal mortality in developmental stages. However, the hatch rate was less than 50% in most of the

backcross progenies. Most of the non-hatching eggs did not contain embryos. The larval siphons of the F₁ hybrids were more like those of *A. sollicitans* than *A. mitchellae*. Hybrid females could be readily distinguished from parental types. No evidence was found for natural hybridization between field populations of *A. sollicitans* and *A. mitchellae*.

Hybridization studies with highly eurygamous mosquitoes must use some type of induced mating technique (McDaniel and Horsfall, 1957). This procedure can sometimes overcome the premating mechanisms that serve to isolate closely related species. The elimination of premating barriers has frequently uncovered additional isolating mechanisms (Davidson *et al.*, 1967; McClelland, 1967; Kitzmiller *et al.*, 1967), including hybrid inviability, hybrid sterility, and hybrid breakdown. Although most hybridization experiments have encountered at least one of these isolating mechanisms, there have been notable exceptions. For example, in the

hybridization of *Aedes sollicitans* and *Aedes nigromaculis* by Fukuda and Woodward (1974), the F₁, F₂ and backcross progeny were fully viable and fertile.

In the present study, we examined the genetic affinity and potential for gene flow between *A. sollicitans* and *A. mitchellae*. Both *A. mitchellae* and *A. nigromaculis* have morphological similarities with *A. sollicitans*. There is a close relationship between larval, as well as adult, characteristics. All three species are highly eurygamous and they can be mated in the laboratory only by induced copulation.

MATERIALS AND METHODS. Field collections of *Aedes mitchellae* were made with

a tractor aspirator (Bidlingmayer and Edman, 1967) in Indian River County, Florida. A few *A. sollicitans* were also obtained in these collections, but many more were obtained from human biting collections in both Florida and Louisiana. The F_1 offspring of these wild caught females were used in the mating experiments.

All mosquitoes were kept in a bioclimatic room with the temperature maintained at 26 ± 1.5 C and the relative humidity at $80 \pm 15\%$. The daily photoperiod was 14.5 hours. Larvae were reared in tap water on the standard diet of O'Meara and Evans (1973). Adults fed *ad libitum* on 10% sugar solution.

Induced mating techniques were modifications of Miura's (1969). The sexes were separated at emergence, even though neither species is known to mate under caged conditions. Females were blood-fed 2 days prior to induced copulation attempts. Mated females were isolated individually in glass shell vials which were lined with moist cheesecloth. Seven days after the eggs were laid, they were submerged in tap water containing a suspension of brewer's yeast (Nutritional Biochemicals Company). All eggs that failed to hatch during a 24-hour period of submergence were examined for embryos by bleaching the egg shell (Mortenson, 1950; Moore and Bickley, 1966).

RESULTS. Viable F_1 hybrids were obtained from reciprocal crosses between *A.*

mitchellae and *A. sollicitans*. In each cross the sex ratio was about 1:1. Hybrid adults appeared normal. Practically no mortality occurred in the larval or pupal stages. The hatch rate was highly variable, but most embryonated eggs hatched when submerged for 24 hours (Table 1).

F_1 hybrids were fertile and they were successfully backcrossed to both parental species. A total of 31 progenies were produced by six different kinds of backcrosses. In general, for each backcross progeny the sex ratio was normal and the larval and pupal mortality was minimal. The hatch rate was less than 50% in most of the backcross progenies. Yet with the exception of a few progenies, most of the unhatched eggs did not contain embryos (Table 2). When the male reproductive system was examined mature, motile spermatozoa were found in nearly all the males from each backcross progeny.

The larvae of *A. sollicitans* and *A. mitchellae* can be best distinguished by morphological characteristics of their siphons. In *A. mitchellae* the siphonal index (ratio of total length to width at base) is usually greater than 3.0 and the pecten teeth extend less than half the length of the siphon, whereas in *A. sollicitans* the siphonal index is less than 2.5 and the pecten teeth extend one half or more of the length of the siphon. The F_1 hybrids had an average siphonal index of 2.4 and the pecten teeth nearly always extended more than half the length of the siphon.

TABLE 1. The occurrence and viability of F_1 offspring from crosses between *Aedes sollicitans* (AS) and *Aedes mitchellae* (AM).

Cross		Female no.	Total no. eggs	Percent hatched	No. non-hatched embryonated eggs	No. 1st instar larvae	No. pupae	No. adults	
♀	♂							♀	♂
AM x AS		1	96	82.3	0	79	78	30	48
		2	48	47.9	6	23	23	16	7
		3	94	50.0	6	47	47	27	20
		4	160	66.9	12	107	105	48	57
		5	87	87.4	6	76	76	40	36
		6	105	69.5	7	73	70	36	31
AS x AM		1	193	12.4	0	24	23	12	9

The siphons of the F_1 hybrid larvae are much more like those of *A. sollicitans* than *A. mitchellae*.

Adults of *A. sollicitans* and *A. mitchellae* differ in several of their scaling patterns. The F_1 hybrids possess some of the characteristics of both parental species. In *A. sollicitans* light and dark scales are scattered over the entire wing, whereas in *A. mitchellae* all wing scales are dark. The wings of the hybrid are covered with both light and dark scales, but the light scales are restricted to the costa and subcosta. The white scaling pattern of the pleural area of the hybrid is intermediate between the dense scaling on *A. sollicitans*

and the rather sparse scaling of *A. mitchellae*. Unlike *A. sollicitans*, most hybrid specimens showed a separation between the scale patches of the sternopleuron and those of the prealar area, but the separation was not as distinct as that found on *A. mitchellae*. Although most hybrids have yellow scaling on the first segment of the hind tarsus, the middle of this segment is seldom completely ringed with yellow scales as it is in *A. sollicitans*. There is no yellow scaling on the legs of *A. mitchellae*. The first abdominal tergite in *A. sollicitans* has a medial patch of yellowish scales, while in both *A. mitchellae* and in the hybrid this medial patch is

TABLE 2. Results of backcrosses of F_1 hybrids to *A. sollicitans* and *A. mitchellae*.

Cross		Female no.	Total no. eggs	Percent hatched	No. non-hatched embryonated eggs	No. 1st instar larvae	No. pupae	No. adults	
♀	♂							♀	♂
AS x F_1 AM/AS	1	226	71.7	2	162	149	71	76	
	2	208	81.3	5	169	77	41	34	
	3	165	1.2	1	2	2	0	2	
	4	192	7.3	0	14	14	7	6	
	5	183	20.8	0	38	37	16	21	
	6	124	37.1	0	46	42	14	27	
	7	197	21.8	1	43	40	18	22	
	8	256	89.1	4	228	178	91	69	
	9	187	77.5	26	145	134	69	61	
	10	221	79.6	31	176	154	84	60	
AS x F_1 AS/AM	1	215	2.3	0	5	5	3	2	
	2	223	45.7	13	102	100	47	36	
	3	167	27.5	0	46	45	19	26	
F_1 AM/AS x AS	1	89	66.3	3	59	56	28	28	
	2	149	21.5	2	32	28	12	16	
	3	135	70.4	10	95	85	43	40	
	4	194	72.2	6	140	101	50	51	
	5	70	70.0	0	49	45	26	19	
	6	95	23.3	40	22	19	8	11	
	7	218	50.5	25	110	90	44	46	
	8	138	1.4	0	2	2	0	2	
	9	67	3.0	0	2	2	0	2	
	10	169	15.4	87	26	26	7	17	
	11	181	3.3	116	6	6	1	5	
F_1 AM/AS x AM	1	149	77.9	4	116	83	44	39	
	2	171	56.1	9	96	68	24	40	
	3	158	23.4	11	37	33	15	18	
	4	123	5.7	2	7	1	0	1	
AM x F_1 AM/AS	1	10	70.0	1	7	5	2	3	
	2	103	30.1	3	31	24	9	14	
F_1 AS/AM x AS	1	153	26.1	30	40	38	19	18	

white scaled. In the hybrid the 5th segment of the mid-tarsus is either entirely dark or ringed with a very narrow band of white scales. In *A. sollicitans* this segment is broadly ringed in white or mostly white, but it is entirely dark in *A. mitchellae*.

DISCUSSION. The results of our crossing experiments indicate a great deal of genetic affinity between *A. sollicitans* and *A. mitchellae*. Numerous F_1 hybrids and backcross individuals were obtained using induced mating procedures (Tables 1 and 2). Moreover, the males of each backcross progeny appeared to develop normal, functional sperm. However, many of the backcrosses produced large numbers of non-embryonated eggs. This situation could have been the result of inadequate sperm transfer. Throughout our study, difficulties were encountered getting the mosquitoes to copulate, even for intraspecific matings. Obviously, other factors might also have precluded embryonic development.

In nature the opportunities for hybridization between *A. sollicitans* and *A. mitchellae* are very limited because the two species occur in different habitats. The larval habitat of *A. sollicitans* is usually the coastal salt marsh or occasionally other areas which contain brackish water, whereas *A. mitchellae* occurs in fresh water. Only in a few areas are both species found in relative close proximity. A light trap located near a salt marsh in Indian River County, Florida has on numerous occasions taken *A. sollicitans* and *A. mitchellae* in the same nightly collection. During the daytime, both species were frequently collected along the same roadside swale with a tractor aspirator. Yet even in these populations we found no evidence of interspecific hybridization such as the occurrence of F_1 hybrids which are distinguishable from either parental species. Apparently, there are other pre-mating isolating barriers in addition to geographical factors which prevent hybridization between *A. sollicitans* and *A. mitchellae*.

Besides providing a means for investigating phylogenetic relationships, experimental hybridization can be utilized to study the genetics of specific morphological, behavioral or physiological characters (Truman and Craig, 1968; Hartberg and Craig, 1973). Sufficient gene flow can be generated by forced mating *A. sollicitans* with either *A. mitchellae* or *A. nigromaculis* to evaluate the degree of dominance associated with certain traits. It is reasonable to assume that the same situation would exist if *A. mitchellae* were crossed with *A. nigromaculis*. In preliminary tests we have successfully hybridized *A. sollicitans* with a third species, *A. taeniorhynchus*. All four species (*A. sollicitans*, *A. taeniorhynchus*, *A. mitchellae*, and *A. nigromaculis*) are definitely closely related, and additional hybridization experiments would surely increase our understanding of the relationships among these mosquitoes.

ACKNOWLEDGMENTS. We thank D. B. Woodard of the Gulf Coast Mosquito Research Laboratory, USDA, for providing us with eggs of *Aedes sollicitans*. The research was supported in part by research grant No. AI-11583 from the National Institutes of Health.

References Cited

- Bidlingmayer, W. L. and J. D. Edman. 1967. Vehicle mounted aspirators. Mosq. News 27: 407-411.
- Davidson, G., H. E. Paterson, M. Coluzzi, G. F. Mason and D. W. Micks. 1967. The *Anopheles gambiae* complex. In: J. W. Wright and R. Pal (ed.). Genetics of insect vectors of disease. Amsterdam: Elsevier Press, pp. 211-250.
- Fukuda, T. and D. B. Woodard. 1974. Hybridization of *Aedes sollicitans* (Walker) and *Aedes nigromaculis* (Ludlow) by induced copulation. Mosq. News 34:71-76.
- Hartberg, W. K. and G. B. Craig, Jr. 1973. Gene-controlled morphological differences in male genitalia of *Aedes aegypti* and *Aedes mascarensis* (Diptera: Culicidae). Mosq. News 33:206-214.
- Kitzmilller, J. B., G. Frizzi and R. H. Baker. 1967. Evolution and speciation within the *maculipennis* complex of the genus *Anopheles*. In: J. W. Wright and R. Pal (ed.). Genetics of insect vectors of disease. Amsterdam: Elsevier Press, pp. 151-210.

- McClelland, G. A. H. 1967. Speciation and evolution in *Aedes*. In: J. W. Wright and R. Pal (ed.) Genetics of insect vectors of disease. Amsterdam: Elsevier Press, pp. 277-311.
- McDaniel, I. N. and W. R. Horsfall. 1957. Induced copulation of aedine mosquitoes. *Science* 125:745.
- Miura, T. 1969. Evaluation of techniques used for mass rearing *Aedes nigromaculis* by induced mating. *Mosq. News* 29:612-616.
- Moore, R. C. and W. E. Bickley. 1966. Hatching of the eggs of *Aedes taeniorhynchus* (Wiedemann) (Diptera: Culicidae) in response to temperature and flooding. *Mosq. News* 26: 405-415.
- Mortenson, E. W. 1950. The use of sodium hypochlorite to study *Aedes nigromaculis* (Ludlow) embryos (Diptera: Culicidae). *Mosq. News* 10:211-212.
- O'Meara, G. F. and D. G. Evans. 1973. Blood-feeding requirements of the mosquito: Geographical variation in *Aedes taeniorhynchus*. *Science* 180:1291-1293.
- Truman, J. W. and G. B. Craig, Jr. 1968. Hybridization between *Aedes hendersoni* and *Aedes triseriatus*. *Ann. Entomol. Soc. Amer.* 61:1020-1025.

XV INTERNATIONAL CONGRESS OF ENTOMOLOGY

FIRST ANNOUNCEMENT

The 15th International Congress of Entomology will be held in the beautiful capital city, Washington, D.C., U.S.A., August 19-27, 1976, under the sponsorship of the National Academy of Sciences and the Entomological Society of America. Sessions will be held in the excellent meeting facilities of the Washington Hilton Hotel. Special events are being planned at national scientific and cultural centers. Two international airports near Washington give direct access from abroad. University housing will be available in addition to hotel facilities.

The Organizing Committee for the Congress is composed of Curtis W. Sabrosky (Chairman and President of the Congress), Ernest C. Bay (Secretary-General), Wallace P. Murdoch (Treasurer), William G. Eden, Gordon E. Guyer, E. F. Knipling, Robert L. Metcalf, John V. Osmun, Ray F. Smith and Edward O. Wilson.

The program will emphasize plenary symposia, invitational speakers, specialized symposia/work groups/panel discussions, and special interest groups or informal conferences. Thirteen program sections cover Systematics, Genetics, Physiology and Biochemistry, Toxicology, Ecology, Behavior, Social Insects and Apiculture, Biological Control, Medical and Veterinary Entomology, Agricultural Entomology and Pest Management, Forest Entomology, Stored Products and Structural Insects, and Pesticide Development, Management and Regulation.

A Congress Brochure and application forms will be mailed in May, 1975. The Brochure will contain information on highlights of the scientific program, receptions, tours, ladies program, scientific, historical and other features of the Washington area, and useful data for visitors.

PLEASE NOTE: Announcements of this Congress are not being sent to individuals, but are being publicized in journals and circulated to museums, departments, and other institutions. If you are interested in receiving future information, including registration forms, please send a postcard to the undersigned with your name and address, typed or in block letters, and also section of your major interest.

DR. ERNEST C. BAY, *Secretary General*, XV International Congress of Entomology, P.O. Box 151, College Park, Md. USA 20740