References Cited

Kenk, R. 1972. Freshwater planarians (Turbellaria) of North America. Biota of freshwater ecosystems identification Manual No. 1. Smithsonian Institution, Washington, D.C. 81 pp.

Legner, E. F. and H. S. Yu. 1975. Larvicidal effects on mosquitoes of substances secreted by the planarian *Dugesia dorotocephala* (Woodworth). Proc. Calif. Mosq. Contr. Assoc. 43:128–31.

Legner, E. F., R. D. Sjogren and I. M. Hall. 1974. The biological control of medically important arthropods. Critical Reviews in Environ. Control 4(1):85–113.

Legner, E. F., R. A. Medved and R. D. Sjogren. 1975. Quantitative water column sampler for insects in shallow aquatic habitats. Proc. Calif. Mosq. Contr. Assoc. 43:110–15.

Legner, E. F., T. C. Tsai, and R. A. Medved. 1976. Environmental stimulants to asexual reproduction in the planarian, *Dugesia dorotocephala* (Woodworth). Entomophaga 21:415–23.

Legner, E. F., H. S. Yu, R. A. Medved and M. E. Badgley, 1975. Mosquito and chironomid midge control by planaria. Calif. Agric.

29(11):3-6.

McConnell, J. V. 1967. On the procuring and the care of planarians. *In*: A Manual of Psychological Experimentation on Planarians, (J. V. McConnell, ed.). Planarian Press, Ann Arbor, Mich. 128 pp.

Medved, R. A. and E. F. Legner. 1974. Feeding and reproduction of the planarian *Dugesia dorotocephala* (Woodworth), in the presence of *Culex peus* Speiser. Environ. Entomol.

3:637-41.

Steel, G. D. and J. H. Torrie. 1960. Principles and Procedures of Statistics. McGraw-Hill Book Co., Inc., New York, 481 pp.

Tsai, S. C. and E. F. Legner. 1977. Exponential growth in culture of the planarian mosquito predator *Dugesia dorotocephala* (Woodworth).

Mosquito News 37 (this number).

Walters, L. L. 1976. Comparative effects of the desert pupfish, Cyprinodon macularius Baird & Girard, and the mosquitofish, Gambusia affinis-affinis (Baird & Girard) on pond ecosystems; and mass rearing feasibility of C. macularius. M. S. Thesis, Univ. of California, Riverside. 249 pp.

Yu, H. S. and E. F. Legner. 1975. Regulation of aquatic Diptera by planaria. Entomophaga

21:3-12.

INHERITANCE OF A NEW MUTANT, PLUM EYE, IN THE MOSQUITO AEDES TOGOI

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ABSTRACT. A new recessive mutant, plum eye (pm), of the mosquito Aedes (Finlaya) togoi expresses dark brown eyes prominently in pupae, though difficulty is encountered in detection of this phenotype in larvae and adults. This allele has been located in the same linkage group as yellow larva (y) and curved wing (c),

which have been tentatively assigned to linkage group 3. The gene sequence and the map units among them were: c - (17-18 units) - y - (40-41 units) - pm. There was positive interference between the two segments, the coincidence coefficient being 0.542-0.636 in the females and 0.685-0.743 in the males.

Six genetic markers thus far known for Aedes (Finlaya) togoi (Theobald), a vector of various species of filariae, have been tentatively assigned to the expected three linkage groups (Tadano, in preparation), since the haploid chromosome number in

this mosquito is three (Suzuki 1939; Sinoto and Suzuki 1943; Rai 1963; Kanda 1968); two of these mutants, yellow larva (y) and curved wing (c), have been placed in linkage group 3, and 16–20 map units exist between the two alleles.

A new mutant plum eye (pm) expresses dark brown eyes most prominently in pupae in contrast to black eyes of wild-type pupae but can be hardly distinguished in larvae and it darkens on emergence. This allele is recessive, highly penetrant, and as viable as the wild phenotype. This paper presents results of the cross experiments to elucidate the linkage relationship among pm, y, and c.

MATERIALS AND METHODS. The yellow larva (y) strain was found in the NGSK strain provided by Department of Medical Zoology, School of Medicine, Nagasaki University, Kyushu (Tadano 1977a). Both curved wing (c) and plum eye (pm) mutants were isolated from the MUR strain collected at Miura City, Kanagawa, Japan, and the MUR strain was utilized as a wild type whenever needed for cross experiments.

Rearing of the mosquitoes and procedures for experiments were the same as those described by Tadano (1976, 1977b). Although mass crosses were done for all experiments, single blood-fed females were isolated into each of plastic cups for oviposition. Three or four days after oviposition, these egg batches were hatched in separate plastic containers in which thereafter the larvae were reared as Classification single families. phenotypes was made in 3rd or 4th instar for y, in pupae for pm, and in adults for sex and c; the proportions of phenotypes including sex in each family were tested by chi2 to see whether the expected ratios (1:1 or 1:3) were obtained. All families in which all the proportions did not show a significant deviation (P>0.05) from the expected ratios were pooled for the subsequent statistical treatment, from which the other families were excluded. Chi2 - tests for genetic linkage were performed according to Bailey (1961) and Serra (1965).

RESULTS AND DISCUSSION. Of 13 crosses undertaken in this study (Table 1), 8 crosses (A through H) are dihybrid experiments and the rest (I through M) are trihybrid crosses; phenotypic scores from dihybrid crosses are tabulated together with those from trihybrid crosses. Four

crosses (A, F, G, and I) were F₁x F₁ and all others were test crosses.

Genetic linkage between the three mutant alleles was examined by the chi2-test for one degree of freedom (table 2). Chi2 values for c y ranged from 82.81 (I) to 431.13 (H), which indicate significantly (P < 0.01) the existence of linkage between the loci. The values for y pm were much greater than 6.635, which is the chi2 value at P = 0.01 for one degree of freedom, though only the value from cross A (5.63) was exceptionally low (0.02>P>0.01). Thus y and pm have been proved to be linked together. The maximum of linkage chi² values for y pm was 65.19 (I), though even a minimum value for cy was 82.81 (I), and so it is very likely that length of the c-y segment is shorter than that of the y-pm segment.

However, the chi² values for c pm were 2.44 or much less and indicate that a free recombination occurred (P > 0.05). Therefore, all the facts mentioned above suggest that the gene sequence is $c ext{-} pm$.

Recombination distances among three loci and the standard errors were calculated for each cross: these are also shown in Table 2, where the standard errors were not estimated from the F1x F1 data. Recombination values between c and y, estimated from testcross data (H, J, K, L, and M), were 15.98 ± 1.41 to $19.53\pm1.18\%$, while the estimation from an F1x F1 cross (I) was 23.24%. On the other hand, recombination units in the y-pm sgement ranged from 34.74 (I) to 42.80±1.73 (J). Thus it is clear that the c-pm segment is so long as to experience a free recombination, as indicated by the recombination value of 47.29 ± 1.73 (L) to 50.14% (G).

Female heterozygotes were employed in crosses C, E, H. K, and M, whereas male heterozygotes were used for crosses B, D, J, and L; from these recombination values no evidence has been obtained that differences in recombination units exist between sexes. Recombination units in the 15.98 ± 1.41 were segment $19.53 \pm \overline{1}.18$ in females, and 16.10 ± 1.27 to 16.85±2.22 in males, the average units females and 18.23 ± 0.92 in being

Table 1. Scores from crosses to elucidate the relationship among $c,\ y,\ {\it and}\ pm$

										Progeny phenotype	henotyp	يو							
		Parental				Fen	Female							Male	le			1	
	age	genotype *)								"	
Cross	Female	e Male	+	Ü	щф	c	8	c	pm y	pm	+	Ü	md	o tpu	2	υp	φm	pm v	Families pooled
A	+ h + + + + + + + + + + + + + + + + + +	$\frac{y}{+pm}$	68	:	37	:	29	:	.c	:	100	:	37	:	44	. :	, o	$\cdot \mid :$	66
я	ypm	$\frac{x}{++}$	196	:	125	:	122	:	208	:	192	:	140	:	145	:	233	:	12
Ö	ypm ++	x ypm	216	:	155	:	126	:	212	:	211	:	151	:	146	:	238	:	10
D	ypm	* ++ x	107	:	82	:	74	:	121	:	144	:	81	:	81	:	118	:	7
ഥ	++ 34m	x ypm	221	:	137	:	168	:	230	;	255	:	169	:	171	:	252	:	∞
H .	$\frac{++}{y b m}$	$\frac{x}{\sqrt{mdy}}$	85	:	23	:	26	:	17	:	96	:	33	:	31	:	21	:	&
Ů	$\frac{ +c }{pm+}$	$\frac{+c}{bm+}$	102	53	43	13	:	:	:	:	137	34	47	20	:	:	:	:	4
Н	+ 20	×	199	41	:	:	46	208	:	:	190	35	:	:	35 2	202	:	:	10
I	$\frac{+ypm}{c++}$	$\frac{x}{c++}$	221	105	58	32	63	ಲ	46	-	231	96	40	73	102	_	99	11	∞
J	c ypm	$\frac{+++}{cypm}$	98	53	93	∞	91	59	25	66	101	16	46	11	7	99	20	93	11
¥	$\frac{+++}{cypm}$	$x \frac{cypm}{cypm}$	125	42	88	10	91	87	57	141	125	33	96	91	11 1	102	30 1	122	6
Г	c ypm	$\mathbf{x} \frac{c y \rho m}{+++}$	96	56	61	14	∞	62	24	93	109	23	93	4	12	73	29 1	102	п
M	c ypm +++	$\frac{cypm}{cypm}$	75	19	28	9	6	70	26	06	65	21	29	20	4	28	18	3 2	7
* Alleles	above the	* Alleles above the line are of maternal origin, those helow the line of naternal origin	naternal	nigin	ţ	lad a	1	1 1 1	90	naternal	ariain.	ĺ							

Alleles above the line are of maternal origin, those below the line of paternal origin.

Table 2. Chi-square values for genetic linkage and recombination values between c, y and pm.

		Chi-square value			Recombination value (%)		
Cross	(Type)	c:y	y:pm	c:pm	c-y	у-т	с-рт
		-	5.63*			39.00	
\mathbf{A}	$(\mathbf{F_1} \times \mathbf{F_1})$					39.09 ± 1.30	
В	(Testcross)		64.81	• • • •		39.73 ± 1.27	
C	(Testcross)		61.44				
Ď	(Testcross)		35.22			39.58 ± 1.73	
	`		61.12			40.24 ± 1.23	
\mathbf{E}	(Testcross)		11.74			39.20	
F	$(\mathbf{F_1} \times \mathbf{F_1})$	• • • • •					50.14
G	$(\mathbf{F_1} \times \mathbf{F_1})$			0.01**			-
H	(Testcross)	431.13			16.42 ± 1.20		
11		82.81	65.19	0.06**	23.24	34.74	49.43
ī	$(\mathbf{F}_1 \times \mathbf{F}_1)$	377.00	16.98	0.59**	16.10 ± 1.27	42.80 ± 1.73	48.66 ± 1.73
J	(Testcross)			0.87**	19.53 + 1.18	38.69 ± 1.48	48.59 ± 1.52
K	(Testcross)	408.94	55.91			39.59 ± 1.70	47.29±1.78
L	(Testcross)	365.34	36.02		16.85 ± 2.22		49.85±1.92
M	(Testcross)	313.02	22.02	0.01**	15.98±1.41	40.98±1.89	49.03 1.32

^{*} 0.02 > P > 0.01.

16.48±0.91 in males. This confirms previous observations (Tadano, in preparation).

Recombination values between y and pm varied from 38.69 ± 1.48 to $40.98\pm1.89\%$ in females, and from 39.09 ± 1.30 to $42.80\pm1.73\%$ in males. The average value was $39.56\pm1.18\%$ in females and $41.18\pm1.21\%$ in males. Therefore, it can be concluded that the two segments, c-y and y-pm, are approximately 17-18 and 40-41 map units, respectively, and that the segment between c and pm is roughly 60 units.

Coincidence coefficients in the *c-pm* segment, calculated from data of test crosses J, K, L, and M, were 0.542 to 0.636 in females and 0.685 to 0.743 in males; they indicate the occurrence of positive interference between the two segments.

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References Cited

Bailey, N. T. J. 1961. Introduction to the mathematical theory of genetic linkage. London: Oxford University Press. 298 pp.

Kanda, T. 1968. Studies on karyotypes of some Japanese mosquitoes. Jap. J. Exp. Med. 38:37-46.

Rai, K. S. 1963. A comparative study of mosquito karyotypes. Ann. Entomol. Soc. Amer. 56:160-170.

Serra, J. A. 1965. Modern genetics. London, New York: Academic Press. 540 pp.

Sinoto, Y. and Suzuki, K. 1943. Karyotypen von einigen Moskitos. Igaku to seibutsugaku 3:175–181.

Suzuki, K. 1939. Chromosomes of mosquitoes (Preliminary note). Jap. J. Genet. 15:296– 298.

Tadano, T. 1976. A new sex-linked mutant, reddish eye, of the mosquito Aedes togoi. Jap. J. Sanit. Zool. 27:247-249.

Tadano, T. 1977a. The inheritance of a mutant yellow larva in the mosquito *Aedes togoi*. Ann. Trop. Med. Parasitol. (in press).

Tadano, T. 1977b. Genetics of three new mutants, straw-colored larva, ruby eye, and pigmented pupa in *Aedes (Finlaya) togoi* (Theobald). J. Med. Ent. (in press).

^{**} Insignificant (P > 0.01).