

rial plates for the counts were made immediately after the spore suspension was prepared when many vegetative, non-insecticidal, cells may have been still viable. In later tests, quantitation plates were prepared just before dosing, and at least 1 day after mixing the bacteria with water.

This strain of *B. thuringiensis* and also *B. sphaericus* have high levels of toxicity to mosquitoes and are seriously considered as biocontrol agents. A comparison of the results here with those of Goldberg & Margalit (1977) indicates for ONR-60A, an even higher level of activity against the simuliids than the culicids, especially when the much shorter exposure times used here are considered.

B. thuringiensis ONR-60A/WHO 1897 appears to offer very real promise for practical application for the control of larval simuliids. When compared with the lepidopterophilic strains of *B. thuringiensis* (Lacey & Mulla 1977) the ONR-60A strain IC₅₀ is at least 1 log lower and the dose response slope much steeper. Continuous exposure of 3rd-5th instar *S. verecundum* larvae to a commercial preparation of *B. thuringiensis* (Dipel[®]) gave an approximate IC₅₀ of 10⁵ spores/ml and increasing the food supply increased the dosage requirement even further (unpublished data).

Because of the spore size and the slow-

ness with which they settle in water the bacterial suspension appears almost ideal as a formulation for a black fly larvicide. Small scale field tests are planned in local streams pending the assessment of pathogenicity towards local stream non-target organisms and the outcome of an application for a permit from the Newfoundland Department of Consumer Affairs and Environment.

References Cited

- Colbo, M. H. and B. H. Thompson. 1978. An efficient technique for laboratory rearing of *Simulium verecundum* S. & J. (Diptera: Simuliidae). Can. J. Zool. 56:507-510.
- Goldberg, L. J. and J. Margalit. 1977. A bacterial spore demonstrating rapid larvicidal activity against *Anopheles sergentii*, *Uranotaenia unguiculata*, *Culex univittatus*, *Aedes aegypti* and *Culex pipiens*. Mosquito News 37:355-358.
- Lacey, L. A. and M. S. Mulla. 1977. Evaluation of *Bacillus thuringiensis* as a biocide of black fly larvae (Diptera: Simuliidae). J. Invertebr. Pathol. 30:46-49.
- Litchfield, J. T. Jr. and F. Wilcoxon. 1949. A simplified method of evaluating dose-effect experiments. J. Pharmacol. Exp. Therapeut. 96:99-113.
- Ramoska, W. A., S. Singer and R. Levy. 1977. Bioassay of three strains of *Bacillus sphaericus* on field-collected mosquito larvae. J. Invertebr. Pathol. 30:151-154.

FAREWELL *fatigans*

Culex quinquefasciatus was described by Say in 1823. *Cx. fatigans* was described by Wiedemann in 1828 and has been recognized as a junior synonym by many specialists for many years. Alan Stone (1956. Proc. Entomol. Soc. Wash. 58:342-343) and Stone, Knight and Starke (1959. Synoptic Catalog . . . Thomas Say Foundation 6:1-258) considered *quinquefasciatus* to be the valid name. However, nomenclatural arguments have persisted. To promote nomenclatural stability, Sunthorn Sirivanakarn and G. B. White have designated a neotype for the southern (tropical) house mosquito (1958. Proc. Entomol. Soc. Wash. 80:360-372). There is no excuse for refusing to use the name *quinquefasciatus*. As T. E. McNeel once said, "If you can't say '*quinquefasciatus*' just call them kinkies." Mosquito workers are indebted to Sirivanakarn and White for a laborious and painstaking task.—W. E. Bickley.