A Pluviometric Fern Spore, Fungal Spore, and Pollen Trap

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ABSTRACT.—Although rain is the most important agent in airborne biological particle deposition most of the current sampling traps retain palynomorphs (fern spore, fungal spore, pollen, among others) but are unable to measure precipitation. The objectives of this study are to present a new simple pluviometric/gravimetric spore/pollen trap and propose a spore/pollen-density rain method, based on the particle frequency and sampling area, which would facilitate ecological inferences about rainfall and Biological Airborne Particle (BAP) deposition, so as to avoid the overrepresentation of the percentage and diverse aerobiological methods. Relative to other spore traps, our proposed trap is simple to build, easy to mount in the field, easy to carry, maintenance free, and requires no energy source. In addition, our trap records rainfall volume, and the quantity of spores captured can be expressed in terms of area-density (particle m^{-2}). The rainfall measured with the trap had no significant differences with the precipitation volume obtained from the pluviometer of the Automatic Meteorological Station at Zacualtipán, Hidalgo, Mexico.

KEY WORDS.—Airborne spore, biological airborne particle, biological particle deposition, efficient spore trap

Aerobiology is a recent scientific discipline that studies the diversity and concentration of biological particles (e.g., pollen, fungal spores, and fern spores) that are transported passively by the atmosphere (Latorre and Caccavari, 2010). Most aerobiological studies focus on the variation of pollen and fungal spore concentrations because of the allergenic effect of these particles (i.e., pollinosis) on humans, and in order to assess the potential for the spread of fungal disease on economic crops (De Benito and Soto, 2001; Lacey and West, 2006). Research has resulted in the development of specialized palynomorph traps, such as the Tauber trap (Tauber, 1974), which is used to analyze pollen deposition (Levetin *et al.*, 2000), and the Burkard

volumetric trap (Hirst, 1952), which was designed to study airborne pollen. Both of these devices are commonly used in many types of research (e.g., Caulton *et al.*, 2000; Hicks, 1999; Kasprzyk, 2004; Yang and Chen, 1998), as are specialized traps, such as the rotorod trap (Murray *et al.*, 2007), the cyclone spore trap (Tate *et al.*, 1980), and others reviewed by Gregory (1961) and Lacey and West (2006).

Although these types of traps can be programmed to survey the air for different periods of time, they are expensive and require energy sources unavailable in some tropical environments (Gupta and Chanda, 1991; Potter and Rowley, 1960). Furthermore, they can be lost or stolen during prolonged field surveys. For these reasons, these traps are mostly used in urban areas, on the rooftops of available buildings (Estrella *et al.*, 2006; Latorre and Caccavari, 2010; Ong *et al.*, 2011; Ting *et al.*, 2010). The use of simpler traps has been reported, such as moss clusters on trees (Limón, 1980), soil samplers (Anupama *et al.*, 2002; Tovar-González, 1987), and exposed petri dishes containing different nutritional media (Brown, 1971). However, simpler traps cannot be used to determine the palynomorph influx, and have low local flora representation (Tejero-Díez *et al.*, 1988). Other traps, such as adhesive slices, have low uptake efficiency in prolonged surveys because of sampling area saturation and particle loss due to rain-washing (Melhem and Makino, 1978). Bush (1992) proposed an inexpensive, phenologically accurate gravimetric palynomorph trap composed of a funnel (sampling area) and a carafe; this trap was later improved by Gosling *et al.* (2003), but both of these traps have the disadvantage of not being able to record pluviometric values. Rainfall is the most important meteorological factor in particle deposition (Ramírez-Trejo, 2002; Ramírez-Trejo *et al.*, 2004; Simabukuro *et al.*, 1998, 2000), yet samplers are unable to measure rainfall.

To preserve the pluviometric values associated with a spore-rain survey and to obtain more ecological inferences about rainfall and palynomorph depositions, we modified and simplified the Bush-Gosling trap.

Our trap consisted of a funnel 9 cm in diameter attached to a two-liter carafe (Fig. 1 A and B). To prevent the evaporation of the rainfall deposited in the trap, a segment of PVC tube 15.5 cm (6.5 in) in diameter was added, surrounding the trap and serving as a base. A mosquito net or another type of mesh could be used to cover the funnel and thus prevent major detritus buildup. Moreover, the cylinder-base top edge should be beveled to avoid over representation of rainfall by splashing.

The uptake area (top of the funnel) can be calculated as $A = \pi r^2$; 63.61725124 cm^2 thus, the total rainfall can be expressed in millimeters by the following equation: $\text{mm} = V/A$, where V is the final volume of rain collected expressed in mm^3 and A is the uptake area expressed in mm^2 . In addition, the particles may be expressed in density (particles/sampling area), which facilitates analysis and avoids the problems related in the percentage method used by Simabukuro *et al.* (2000).

The advantage of this trap is that it allows better aeropalynological interpretations with the best airborne particle deposition factor (rain). To test

FIG. 1. Pluviometric spore/pollen trap. A. Pluviometric spore/pollen trap diagram: 1) funnel, 2) screen mesh, 3) beveled top edge PVC tube, 4) security seal to secure the mesh, 5) recapping with a hole for the funnel tube, 6) carafe with a 2 L capacity, 7) wall of the PVC tube. B. Trap mounted at an experiment site. C. Measuring and filtering of the trap content with different sized mesh: 1) 100 µm mesh, 2) 20 µm mesh, 3) graduated cylinder. D. Details of simple filter build up by a PVC tube segment, with a 100 µm mesh attached with a clamp. E. Particles captures with the device at 10 × magnification. F. Striate trilete fern spore aff. *Alsophila firma* (Baker) D. S. Conant (arrow), surrounded by diverse pollen grains at 100 × magnification.

durability and efficiency of the trap, we used it to determine the spore rain in an area near the Malila River in the state of Hidalgo, Mexico. The trap was left in the field in its cylinder-base with monthly trap changes, which did not result in damage from environmental conditions.

To vary the sampling period, one can change the carafe capacity and funnel size according to the total rainfall observed in previous years. Because we

TABLE 1. Total number of fern spores captured by six traps, mean precipitation, and spore density in a monthly one-year survey (March 2009 to February 2010). PST =Pluviometric Spore Trap.

Period	Total fern spore counted	PSTM mean precipitation (mm)	Fern spore density (spore m ⁻²)
March 2009	621	10.58	16269.2
April 2009	335	10.27	8776.4
May 2009	228	73.09	5973.2
June 2009	274	115.66	7178.3
July 2009	123	7.86	3222.4
August 2009	224	226.33	5868.4
September 2009	129	204.61	3379.5
October 2009	83	154.31	2174.4
November 2009	66	55.80	1729.1
December 2009	101	47.42	2646.0
January 2010	149	98.50	3903.5
February 2010	129	24.49	3379.5

knew the maximum precipitation rates in our study site, we selected a volume capacity of two liters in order to avoid overflow.

With this trap, the palynomorphs and rain remained in the carafe and were later separated by filtration with different size mesh in the laboratory (Fig. 1 C and D). One hundred µm mesh was used to remove medium detritus and 20 µm mesh was used to collect the particles of interest. In addition, rainfall volume was measured.

The 20 µm mesh was rinsed with 70% ethanol, and the ethanol was collected in a 15 ml vial. The liquid was then centrifuged at 1500 rpm for 3 minutes. The resulting precipitate was then suspended with 5 ml of 70% ethanol from which semi-permanent preparations were made.

FIG. 2. Monthly precipitation values (mm) of the Automatic Meteorological station at Zacualtipán, Hidalgo, México (AMS Zac) and the mean values of six pluviometric spore/pollen traps (PST) localized at 14 km of AMS Zac, from March 2009 to February 2010.

After a one-year test, the trap showed the ability to capture diverse biological particles such as pollen, fungal spores, and fern spores (Fig. 1 E, F), the latter of which are shown in Table 1. Also the comparison between the precipitation values obtained with six trap devices and those obtained by the pluviometer of the nearest weather station, using a Mann-Whitney U-test, showed no significant differences ($W=52$, $P=0.51$, $\alpha=0.05$, Fig. 2).

Spore perine characters offer more accuracy in the identification of fern taxa and therefore we recommend using a fresh mount rather than the common Erdtman (1960) acetolysis method, in order to preserve the fragile perine of some taxa (Devi, 1980).

This simple trap could help with the determination of phenological processes in tropical environments and thus address future challenges, including understanding the possible consequences of global warming on ferns and their spore dispersal (Mehltreter, 2008).

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