Chapter 5

Herpetofauna of Montane Areas of Tanzania. 1. Results from Two Amphibian Surveys of Malundwe Mountain, Mikumi National Park

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Abstract

As part of a larger survey of flora and fauna of Malundwe, funded by Critical Ecosystems Partnership Fund, amphibians were collected on three separate surveys of this mountain between 2004 and 2007. These surveys constitute the first surveys for amphibians in this part of the Eastern Arc Mountains, a global biodiversity hotspot. Samples from these surveys revealed an amphibian fauna with links to the rest of the Eastern Arc Mountains. Only relatively few species were found during these surveys, probably due to a small cumulative sampling effort, lack of a permanent water source, and the very small size of this forest fragment. The amphibian fauna of Malundwe shares an affinity with the Uluguru Mountains, both in terms of species composition of each mountain block and genetic proximity for species with available population level data. I describe the species and associated microhabitat from the available voucher specimens resulting from the Malundwe surveys.

Introduction

Malundwe Mountain is a little-known and often overlooked massif of the Eastern Arc Mountain (EAM) chain in Tanzania. With a total forested area of only 4.5 km², this section of afromontane rainforest is one of the smallest blocks in the chain, and one of only two protected by Tanzania National Parks (TANAPA) (Lovett & Norton, 1989; Stanley et al., 2005). Recent surveys, funded through the Critical Ecosystems Partnership Fund (CEPF), have helped to document the floral and faunal diversity of this series of peaks and to explore the relationship to other mountain ranges in close proximity (e.g., the Uluguru and Udzungwa Mountains). This study constitutes the first survey of amphibians in this Eastern Arc Mountain fragment and, given the pattern of endemism in the region (Poynton et al., 2007; Blackburn, 2009; Blackburn & Measey, 2009), is an important contribution to our understanding of amphibians in this global biodiversity hotspot (Myers et al., 2000).

Study Area

Malundwe Mountain is located approximately 300 km west of Dar es Salaam in southern Mikumi National Park (MNP) (7°24'S, 37°18'E; Fig. 1) in central/eastern Tanzania. The three peaks that make up this small range reach a maximum height of 1250 m elevation, with afromontane forests extending from the peak down to 800–900 m on the eastern

slopes (Lovett & Norton, 1989). The total amount of forested area, divided between two patches, is 4.5 km², with a closed forest canopy height of 20–30 m (Lovett & Norton, 1989). Along with the Taita Hills, this total forest area comprises one of the smallest patches of forest in the entire Eastern Arc (Newmark, 2002; Burgess et al., 2007). Malundwe is unique, however, in that the entire area of mid-elevation forest is naturally very small (unlike the Taita Hills, where deforestation is a major factor in the small area of extant forest) and has been isolated and protected in the middle of a large area of preserved savannah, likely never affected by human encroachment. There is only one source of groundwater among these peaks: a small stream flowing adjacent to the research camp area established for biodiversity surveys (Fig. 1) (7°23.719'S, 37°18.279'E, 985 m) (Stanley et al., 2007). This stream can also be encountered farther down the slope as a slow-moving stream/wetland near the base camp (7°21.913'S, 37°17.338'E, 840 m) (Stanley et al., 2007). The water source is not permanent and is often completely dry by August until the rainy season beginning in December (observation by Animal Behaviour Research Unit research team (ABRU), Mikumi National Park).

Surveys

The amphibians collected during survey work in 2007 were incidentally encountered during small mammal surveys through digging of pitfall lines (see Chapter 1 of this volume),

FIG. 1. Maps of the Eastern Arc Mountains with Malundwe boxed (a) and in Mikumi National Park in Tanzania (b); elevational map of Malundwe peak showing research camps (c). Figure created by Rebecca Banasiak and used with permission from Stanley et al. 2005.

collected in pitfall lines, and through chance encounters while walking. An additional incidental survey took place in 2004, but the material appears to have been lost. In contrast, in 2006, an effort was made to survey specifically for the herpetofauna of this region by including a herpetologist (Lawson) in the survey team. The 2005 survey was conducted in July (dry season) over a 10-day period (2005), with no concentrated effort collecting amphibians. The survey in 2006 was conducted over a seven-day period in early May (rainy season) with a distinct goal of cataloguing amphibians. The survey team followed the route of Stanley et al. (2007) to reach the Malundwe research camp with one night at the beginning of the survey and one night at the end of the survey at a base camp at the edge of the forested peaks ($7^{\circ}21.913'S$, $37^{\circ}17.338'E$, 840 m), where we searched for amphibians in the evenings. We spent six nights at the research camp searching for amphibians night and day ($7^{\circ}23.719'S$, $37^{\circ}18.279'E$, 985 m).

Methods

During the 2005 survey, caecilians and frogs were collected that were either recovered while installing pitfall buckets, caught in pitfall lines, or encountered while walking through the forest during the day. Herpetological surveys in 2006 were conducted primarily in areas within 200 m of water and extending along crests in the hills through pitfall lines. Each day, researchers searched for frogs among the leaf litter, on trees or other vegetation, and in the pitfall buckets. At night, researchers again searched leaf litter, trees, and low vegetation near water sources. Whenever possible, call recordings were made during the 2006 foray with a Marantz PMD660 Digital Recorder and linked to the associated specimen (FMNH collection). For processing specimens, each individual was first photographed and euthanized with MS222. Tissues were then taken from the liver and inner thigh and stored in DMSO buffer for molecular analysis. Specimens were then preserved

in formalin (10%), rinsed in water, and stored in ethanol (70%) for morphological analysis. Standard external measurements were taken for each specimen. Each specimen was assigned a field number based on the collector (Leah C. Collett, LCC; Lucinda Lawson, LL; Kate B. McQuaid, KBM; David C. Moyer, DCM; William T. Stanley, WTS). Specimens are currently deposited at the Field Museum of Natural History, Chicago (FMNH); the Natural History Museum, London (BMNH); and the Museo Tridentino di Scienze Naturali, Trento, Italy (MTSN). Specimens listed as "cf." indicate that the taxon shares an affinity with a known species, but slight morphological variation questions a definitive ID. Further morphological and molecular investigations are being conducted, and full species descriptions will be published following these studies.

Results

The incidental amphibian encounters of 2005 and 2007 yielded 13 anurans and two caecilian specimens, all from the research camp area (higher elevation). The species found in this survey are: *Arthroleptis* cf. *affinis*, *A. stenodactylus*, *A. xenodactyloides*, *Probreviceps* cf. *macrodactylus*, *Scolecomorphus* cf. *vittatus*, and *Boulengerula* cf. *uluguruensis*.

The week-long survey of 2006 encountered a total of 34 amphibians. At the base camp (lower elevation), we encountered *Afraxalus fornasini*, *A. stuhlmanni*, *Arthroleptis stenodactylus*, *Hyperolius puncticulatus*, *Ptychadena oxyrhynchus*, and *Ptychadena* sp. At the research camp, we encountered *Arthroleptis* cf. *affinis*, *A. stenodactylus*, *A. xenodactyloides*, *Leptopelis uluguruensis*, *Nectophrynoides* cf. *tornieri*, *Boulengerula* cf. *uluguruensis*, and *Scolecomorphus* cf. *vittatus*.

Annotated List of Amphibians Collected from Malundwe Mountain

Voucher specimens are referred to by Museum accession numbers where available and field numbers (e.g., WTS, LCC) for those yet to be accessioned at the Natural History Museum (NHM), London.

Anurans

Afraxalus fornasini (Bianconi, 1849)

The individual found possesses the dorsal color pattern of broad light bands from the nostrils to the urostyle separated by a thick, dark central band. Sides and feet are dark brown. The specimen is a large female with large, conspicuous, black-tipped asperities on head, back, and the dorsal surfaces of limbs. These characteristics, along with the tibia/snout–urostyle length (SUL) ratio of this specimen (45%), correspond with Pickersgill's (2007) description. The head width/tibia ratio, however, is much higher than his description (106% instead of the 51%–80% ratio described). The dorsal pattern of this individual clearly most closely resembles photos depicting *A. fornasini* as opposed to *A. unicolor* in Pickersgill (2007), which Pickersgill considers to be potentially a separate species. Further molecular work will be necessary to determine whether these are different

species and, if so, which of the two species this individual belongs to (Poynton, 1964; Schiøtz, 1975; Poynton & Broadley, 1987b; Schiøtz, 1999).

SPECIMENS EXAMINED—FMNH 275600.

Afraxalus stuhlmanni (Pfeffer, 1893)

Individuals have the typical dorsal pattern described for this species, a pair of slightly convergent dark lines from the rump over the back to between the eyes, with a scattered distribution of asperities (Schiøtz, 1974; Poynton & Broadley, 1987a; Pickersgill, 2005; Poynton, 2006).

SPECIMENS EXAMINED—FMNH 275602–275607.

Arthroleptis cf. *affinis* (Ahl, 1939)

Based on morphological measurements, the individuals from this survey fall well within the characteristics of *A. affinis* outlined by Poynton and Loader (2008). Head width/SUL ratios for individuals from this collection ranged from 38.3%–45.2%, within the ranges (34.8%–45.9%) given by Poynton and Loader (2008). Additionally, inner metatarsal tubercle/head width ratios ranged from 11.3%–14.2%, most of which were in the known range of *A. affinis* of 8.70%–13.77%. The highest ratio (14.2%) is from a poorly preserved specimen (making it difficult to measure the tubercle), and if excluded, these frogs have a range from 11.3%–12.6%. Molecular data show this population to be closely associated to Uluguru populations (Channing, 2001; Channing & Howell, 2006; Blackburn, 2008; Poynton & Loader, 2008).

SPECIMENS EXAMINED—FMNH 272056; WTS 6768, 6795, 6820, 6832; LCC 009.

Arthroleptis stenodactylus (Pfeffer 1893)

Specimens have the heavily built body with relatively short rear legs characteristic of this species (Poynton & Loader, 2008). Inner metatarsal tubercles are distinct and shovel shaped (Poynton, 1964; Poynton & Broadley, 1985; Channing, 2001; Channing & Howell, 2006).

SPECIMENS EXAMINED—FMNH 272054, 272055, 275599; LCC 011, 013; WTS 6850.

Arthroleptis xenodactyloides (Hewitt 1933)

Adults from this collection range in size from 12.95 to 19.10 mm. Dorsal coloration on some individuals is mottled brown with banded arms and legs. Ventral surface of limbs and throat are gray/pink. Ventral surface is white/gold. Digits have small distinct discs in a teardrop shape, and hands and feet are marked with many tubercles (Poynton, 1964; Poynton & Broadley, 1985; Channing, 2001; Channing & Howell, 2006).

SPECIMENS EXAMINED—WTS 6802, 6764–6756; FMNH 272057.

Hyperolius puncticulatus (Pfeffer 1893)

Although the pattern in *H. puncticulatus* can be variable, the Malundwe population appears solely to exhibit the most common color morph: yellow dorsum with conspicuous black-lined canthal and dorsolateral stripes (Schiøtz, 1975; Poynton & Broadley, 1987c; Schiøtz, 1999).

SPECIMENS EXAMINED—FMNH 274351–274359.

***Leptopelis uluguruensis* (Barbour and Loveridge 1928)**

Dorsal coloration of these individuals ranged from a brown-green background at night to blue-green during the day, mottled with small yellow spots. Individuals possess red-gold pupils/iris. The ventral surface is white with very blue points at shoulder joints, waist, and knee and ankle joints. The heads are characteristic of the species, with a broad shape and short nose (Schlötz, 1975, 1999).

SPECIMENS EXAMINED—FMNH 275626–275628.

***Nectophrynoides cf. tornieri* (Roux 1906)**

The dorsal coloration of this individual is mottled black/brown, with dark sides. Legs, feet, toes, and thighs are speckled. Ventral side is dark and flecked with white dots. The throat of this individual is pale grey with white flecks. Groin and toes are pink. Eyes are golden. The head is thick at the base (spade shaped), and the back and legs are covered in small warts (Roux, 1906; Channing, 2001; Menegon et al., 2004; Channing & Howell, 2006). The taxonomic status of this individual is currently being assessed by M. Menegon and S. P. Loader.

SPECIMENS EXAMINED—MTSN 7329.

***Probreviceps cf. macrodactylus* (Nieden, 1926)**

This is a juvenile specimen in which a tympanum is visible. Juveniles of the subspecies *P. macrodactylus* and *P. loveridgei* are difficult to distinguish; thus, further surveys are needed.

SPECIMENS EXAMINED—LCC 010.

***Ptychadena oxyrhynchus* (Smith 1849)**

Although this specimen (and potentially the unknown *Ptychedena* sp. listed below) appears to have not reached adult size, it is identifiable based on morphological characteristics outlined in Channing (2001), Channing and Howell (2006), and Pickersgill (2007). This specimen has several dorsolateral ridges, and as fitting with the description of the species, the two median ridges begin at the posterior of the eye and end in the middle of the back. Distal to these medial ridges, two other ridges extend from behind the eyes and continue to the vent. The tympanum (typ) is clearly visible, and it reaches almost the eye diameter (typ/eye = 0.88, in line with the described range of 0.8–1.0). The hind legs are long and muscular as described for the species; the ratio of femur/SUL is 57%. The tibia/SUL ratio is also slightly small, 63% as opposed to 0.7–0.8 (Channing 2001), or 67%–77% (Pickersgill 2007). Foot and longest toe measurements, however, are slightly high (0.92) for the range (0.8–0.9). Toe-tips and finger-tips are not enlarged. Eye diameter = 4.69 mm, tympanum width = 4.12 mm (tympanum height = 3.5 mm). Femur = 22.2 mm, tibia = 24.6 mm, foot = 35.77 mm, inner metatarsal = 3.00 mm. The distance from the nostrils to the snout is approximately equal to the internarial distance. This specimen is dark green/brown with gold ridges down the back. Speckled brown/cream sides. The upper section of the iris is golden, throat and dorsal surfaces white. Underside of thighs pinkish with cream spots. Back of thigh had light and dark longitudinal bands (Schmidt & Inger, 1959; Poynton, 1964; Inger, 1968; Channing, 2001; Channing & Howell, 2006; Pickersgill, 2007).

SPECIMENS EXAMINED—FMNH 275593.

A second possible *P. oxyrhynchus* specimen was also found, although as a juvenile, the identity is uncertain. The specimen

has a long snout and three horizontal bands on the back of the thighs connecting the knees.

SPECIMENS EXAMINED—FMNH 275601.

Gymnophiona

***Boulengerula cf. uluguruensis* (Barbour & Loveridge 1935)**

The specimens closely match the description of *Boulengerula uluguruensis* by Nussbaum and Hinkel (1994), particularly in the coloration and annuli counts. Furthermore, Loader et al. (in press) indicate a close phylogenetic relationship between Uluguru populations of *B. uluguruensis* to Malundwe, although there is substantial genetic difference between these populations.

SPECIMENS EXAMINED—LCC 003; KBM 003. These are in the process of being accessioned into the BMNH collection.

***Scolecophorus cf. vittatus* (Boulenger 1895)**

Three of the four specimens found for this species have annuli counts within the regular range for this species (130, 133, and 136). The fourth specimen (KBM 005) extends the variation of number annuli for this species to 150 (148 was the previous maximum known). The coloration of the specimens resembles that of Nussbaum's (1985) description of *S. vittatus* (Loader et al., 2003a,b; Gower et al., 2004).

SPECIMENS EXAMINED—LCC 004; KBM 002, 004, 005. These are in the process of being accessioned into the BMNH collection.

Discussion

Although this is only the first, and relatively preliminary, investigation of the amphibians of Malundwe, the collection offers a glimpse of the diversity in this poorly known Eastern Arc forest fragment. The small isolated peaks of the Malundwe range contain a small subset of the known amphibian diversity of larger Eastern Arc Mountains (Table 1). This reduced biodiversity may be a result of the small size of this fragment, which is a fraction of the size of most others in this system. For example, Malundwe has a total forested area of 4.5 km², whereas the other mountains, although heavily deforested in many cases, are hundreds of square kilometers. Pare Mountains have 484 km² of forest, West Usambara 328 km², East Usambara 413 km², Nguru 647 km² including Nguu, Ukaguru 184 km², Uluguru 527 km², Rubeho 499 km², Udzungwa 1960 km², and Mahenge 291 km² (Newmark, 1998). The Taita Hills are the only comparable area with just 6 km² of forest, although the original amount was much greater. In comparing Malundwe species diversity to lists from the Udzungwa, Uluguru (Malundwe's nearest neighbor), and the East Usambara Mountains, it is apparent that the species encountered at Malundwe appear to be the omnipresent types, found on many mountain areas throughout the EAM. When species are shared between a subset of mountains and Malundwe, those on Malundwe appear to be more similar to the northern localities (the Uluguru Mountains and the East Usambara) than the southern locality (Udzungwa). Based on molecular datasets, preliminary work on *Boulengerula uluguruensis* and *Hyperolius puncticulatus* suggests that Malundwe populations show the closest phylo-

TABLE 1. Comparison of amphibians encountered on Malundwe Mountain and lists compiled from the literature and chapters in this edited volume from the East Usambara, Uluguru, and Udzungwa Mountains.

	Udzungwa	Malundwe	East Usambara	Uluguru
Anura				
<i>Afrixalus brachycnemis</i>	X		X	X
<i>Afrixalus delicatus</i>	X			X
<i>Afrixalus fornasini</i>	X	X	X	X
<i>Afrixalus morerei</i>	X			
<i>Afrixalus stuhlmanni</i>		X	X	X
<i>Afrixalus uluguruensis</i>	X		X	X
<i>Amietia angolensis</i>	X		X	X
<i>Amietophrynus gutturalis</i>	X		X	X
<i>Amietophrynus maculatus</i>	X		X	X
<i>Arthroleptides martiensseni</i>			X	
<i>Arthroleptis affinis</i>	X	X	X	X
<i>Arthroleptis reichei</i>	X			
<i>Arthroleptis stenodactylus</i>	X	X	X	X
<i>Arthroleptis xenodactyloides</i>	X	X	X	X
<i>Arthroleptis xenodactylus</i>			X	
<i>Arthroleptides yakusini</i>	X			X
<i>Breviceps mossambicus</i>	X		X	
<i>Bufo brauni</i>	X		X	X
<i>Bufo lindneri</i>	X			
<i>Callulina krefftii</i>	X		X	X
<i>Chiromantis xerampelina</i>	X		X	X
<i>Hemissus marmoratus</i>	X		X	
<i>Hildebrandtia ornata</i>	X			
<i>Hoplophryne rogersi</i>			X	
<i>Hoplophryne uluguruensis</i>	X			X
<i>Hyperolius argus</i>			X	
<i>Hyperolius kihangensis</i>	X			
<i>Hyperolius minutissimus</i>	X			
<i>Hyperolius mitchelli</i>	X		X	X
<i>Hyperolius parkeri</i>	X		X	
<i>Hyperolius pictus</i>	X			
<i>Hyperolius puncticulatus</i>	X	X	X	X
<i>Hyperolius spinigularis</i>			X	X
<i>Hyperolius tuberilinguis</i>	X		X	
<i>Hyperolius mariae</i>			X	
<i>Kassina maculata</i>	X		X	
<i>Kassina senegalensis</i>	X		X	
<i>Leptopelis barbouri</i>	X		X	
<i>Leptopelis flavomaculatus</i>	X		X	X
<i>Leptopelis parkeri</i>	X		X	X
<i>Leptopelis uluguruensis</i>	X	X	X	X
<i>Leptopelis vermiculatus</i>	X		X	
<i>Mertensophryne micranotis</i>			X	X
<i>Nectophrynoides asperginis</i>	X			
<i>Nectophrynoides cryptus</i>				X
<i>Nectophrynoides frontierei</i>			X	
<i>Nectophrynoides laevis</i>				X
<i>Nectophrynoides minutus</i>				X
<i>Nectophrynoides poyntoni</i>	X			
<i>Nectophrynoides pseudotornieri</i>				X
<i>Nectophrynoides tornieri</i>	X	X	X	X
<i>Nectophrynoides viviparus</i>	X			X
<i>Nectophrynoides wendyae</i>	X			
<i>Parhoplophryne usambarica</i>			X	
<i>Phrynobatrachus acridoides</i>	X		X	X
<i>Phrynobatrachus krefftii</i>			X	
<i>Phrynobatrachus mababiensis</i>	X			
<i>Phrynobatrachus natalensis</i>	X			X
<i>Phrynobatrachus parvulus</i>	X			
<i>Phrynobatrachus rungwenensis</i>	X			
<i>Phrynobatrachus uzungwenensis</i>	X			X
<i>Probreviceps macrodactylus</i>	X		X	X
<i>Probreviceps rungwenensis</i>	X			
<i>Probreviceps uluguruensis</i>				X
<i>Ptychadena anchietae</i>	X		X	
<i>Ptychadena grandisonae</i>	X			
<i>Ptychadena mascareniensis</i>			X	
<i>Ptychadena mossambica</i>	X			
<i>Ptychadena oxyrhynchus</i>		X	X	X
<i>Ptychadena porosissima</i>	X			

TABLE 1. Continued.

	Udzungwa	Malundwe	East Usambara	Uluguru
<i>Ptychadena taenioscelis</i>	x			
<i>Ptychadena uzungwensis</i>	x			
<i>Pyxicephalus edulis</i>	x			
<i>Schismaderma carens</i>	x		x	
<i>Spelaophryne methneri</i>	x			x
<i>Stephopaedes usambarae</i>			x	
<i>Strongylopus fuelleborni</i>	x			
<i>Xenopus laevis</i>	x		x	x
<i>Xenopus mulleri</i>			x	
<i>Xenopus cf. petersii</i>	x			
Gymnophiona				
<i>Scolecormorphus vittatus</i>		x	x	x
<i>Boulengerula ulugurensis</i>		x		x
<i>Boulengerula boulengeri</i>			x	
<i>Scolecormorphus kirkii</i>	x			x
<i>Scolecormorphus uluguruensis</i>				x

genetic relationships to Uluguru populations (Lawson, 2010; Loader et al., in press). The similarity of the overall amphibian communities and the low genetic diversity between populations of species on Malundwe and the Uluguru Mountains indicate that these forest fragments are among some of the more closely related but separated areas within the Eastern Arc system (Loader et al., 2004; Poynton et al., 2007; M. Menegon, unpubl. data). Unfortunately, species lists throughout the Eastern Arc are too incomplete and mountain ranges too unevenly sampled to make broad comparisons of assemblages (Poynton et al., 2007).

The Malundwe block is one of the smallest of the Eastern Arc blocks and has been found to be completely without water during the dry season (L. Collett, pers. com.: dry season foray by ABRU team, 2004). Both the small total forest area and the potential vulnerability of the area to drought might mean that amphibian populations in Malundwe may not be stable through time, possibly acting as a metapopulation sink with the genetically and spatially proximal Uluguru Mountains. The description of the Malundwe Mountain amphibians fills a gap in our understanding of endemism and species distributions throughout the Eastern Arc, addressing connectivity between northern and southern mountain populations. With focused sampling effort, the biogeographical connections of Malundwe with the other Eastern Arc fragments and the distinctiveness of its amphibian assemblage will aid biologists and conservationists in understanding the diversity within this dynamic area. Although, at present, Malundwe appears to only have widespread species, a closer examination may reveal some of those to be specifically distinct from other EAM species and endemic to Malundwe.

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